The Role of Microbes on the Fate of PFAS in the Environment

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EGLE

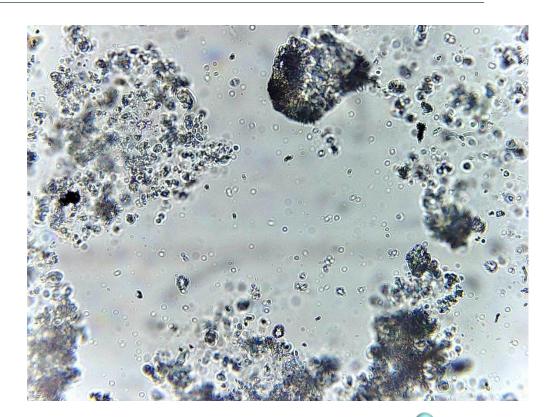
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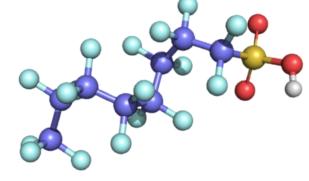
October 12, 2023

Outline

- Per and polyfluorinated alkyl substances (PFAS)
 - Background
 - Biotransformation
- Endemic Microbes Alpena Hide and Leather (AHL)
 - Benchtop 1.0
 - Benchtop 2.0
 - Field Results





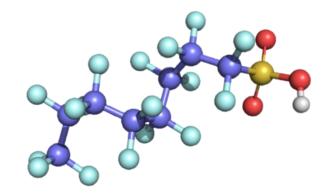




Acknowledgments

- EGLE Janice Adams and Mike Jury
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- Fixed Earth Tim Repas
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- Bureau Veritas Laboratory Heather- Lynn Lord and Lori DeFour







The Role of Microbes on the Fate of PFAS in the Environment

PFAS – General Site Introduction Biotransformation – The gift that keeps on giving



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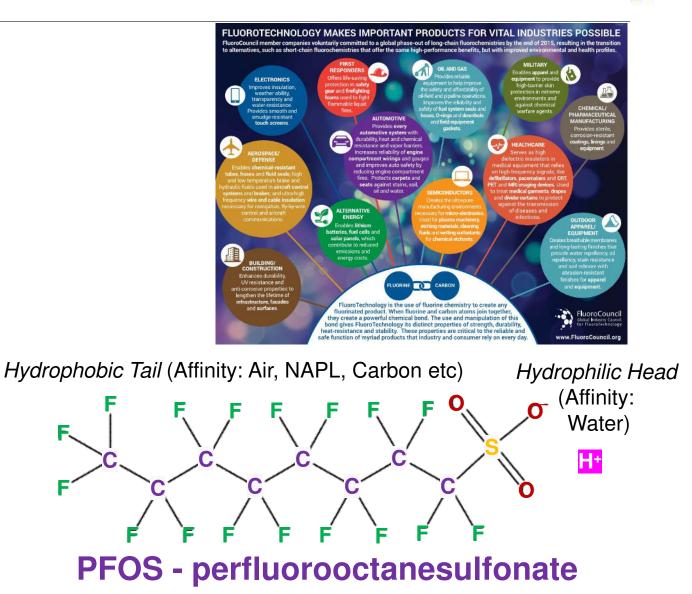
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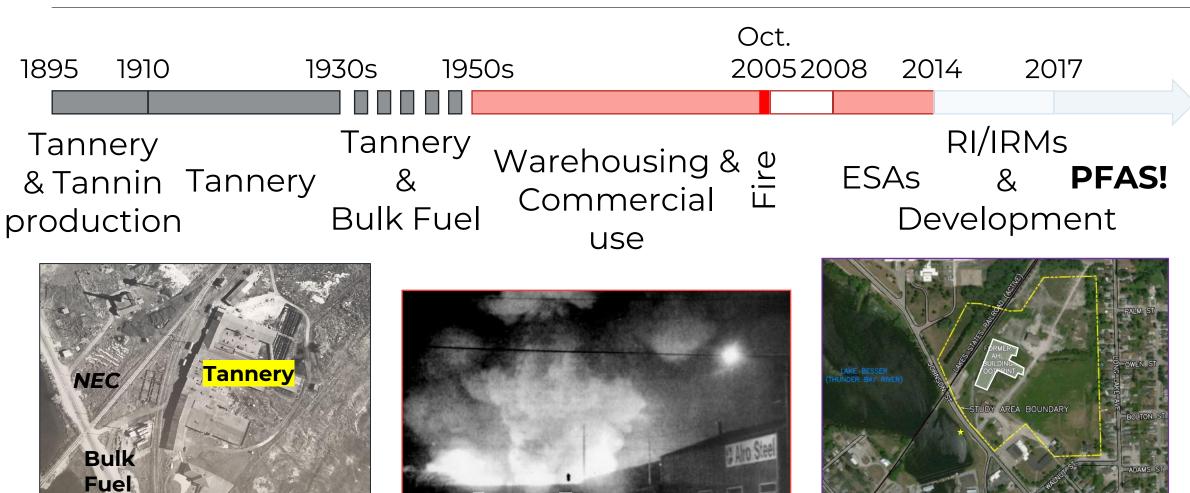
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PFAS – Emerging Contaminants of Concern

- Family of widely used compounds
- Strong carbon-fluorine bond
- Persistent in environment "Forever Chemicals"
- Surfactants with hydrophobic "tails" and hydrophilic "heads"
- Cationic (+), anionic (-), or zwitterionic (+ and -)
- Mechanisms to cleave the "head" debones the snake (Trang et al. 2022)

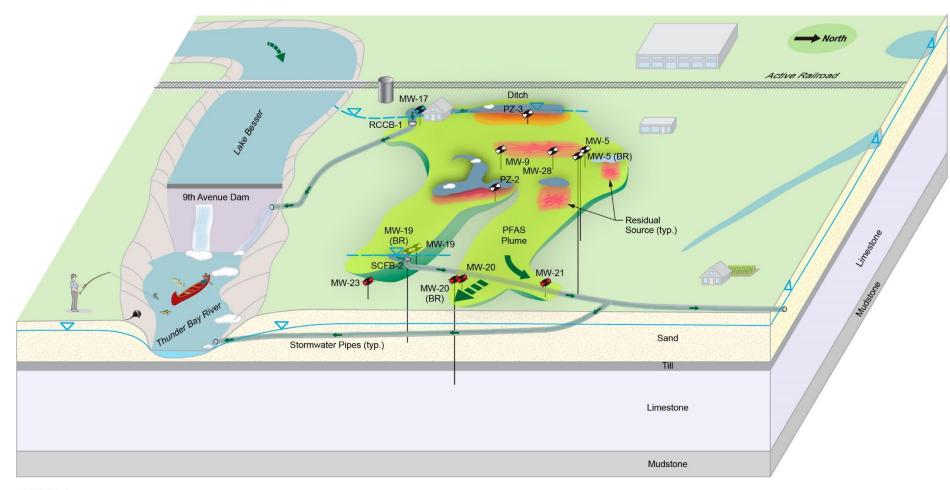




Former Tannery Building



Alpena Hide and Leather – Conceptual Site Model

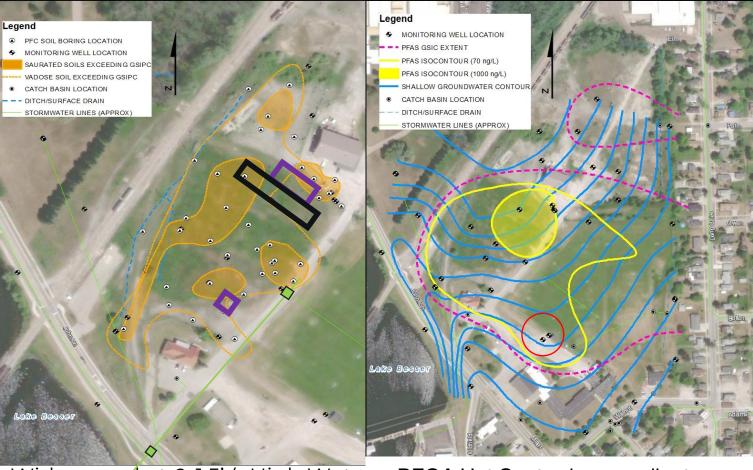


Not To Scale

PFAS in Soil & Groundwater

- Source: Surface to Capillary Fringe (low) - Migration to storm water?
- Soil to Groundwater: Detections "not 1:1" - "Flushing" fractionation
 - Precursor transformation
 - Limits of detection?

PFAS	Soil		Groundwater		
	ng/g (ppb)	f (%)	ng/L (ppt)	f (%)	
PFBA	ND	0%	493	92%	
PFBS	5.7	11%	3,140	85%	
PFHxS	43	56%	15,400	79%	
PFOS	264	63%	8,270	74%	
PFOA	5.4	9%	804	79%	
Total (537)	14 – 76% samples		20 – 92% samples		



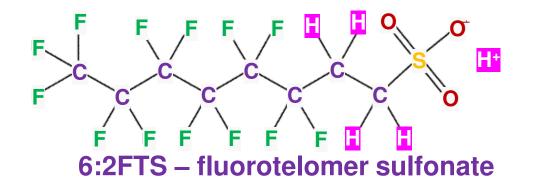
- —Widespread at 0-1.5' (~High Wate PFOA Hot Spot downgradient
- -Hotspots at 4-5 feet (~Low Water) Expanding plume - Former building footprint
 - Topographic lows/infiltration areas
- Limited detections at base of aquifer

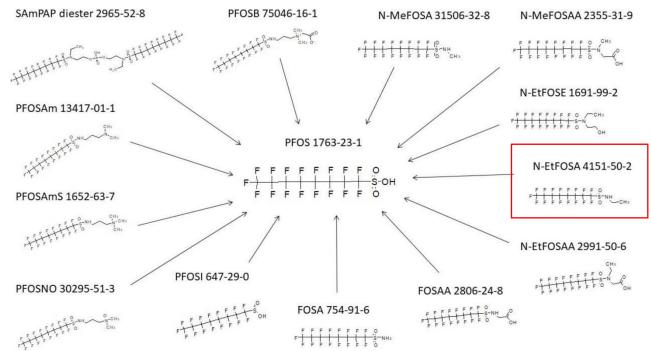
- Offsite fractionation: PFBA+PFBS, followed by PFOA then PFOS



Polyfluorinated Alkyls

- Poly-fluoros: "precursors" that can transform to per-fluoros
- Bio-transformation of poly- to perfluoro alkyls has been demonstrated:
 - Anaerobic (e.g., Yi et. Al. 2014)
 - Aerobic (e.g., Zhang et al., 2020)
 - Fungal (e.g., Shah et al. 2023)
- Bio-transformation has been shown to be a "team" sport
- Inorganic inhibitors (e.g., sulfates) can shut down some transformation
 reactions (Yang et al, 2023)

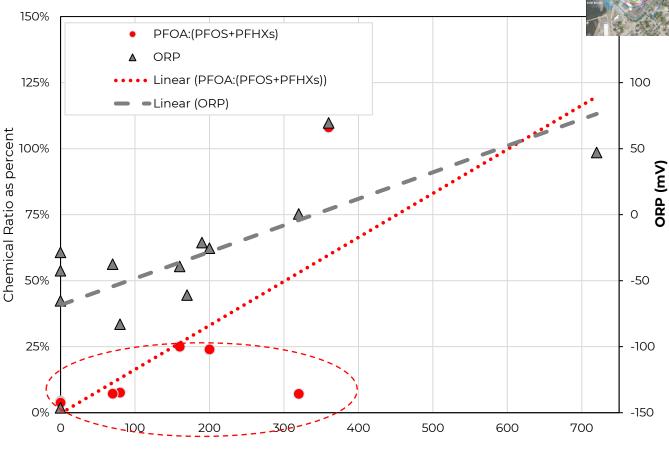




Precursor Compounds – PFOA

- PFOA increases in concentration **off-site**
- PFOA increases relative to PFHxS and PFOS
- Formation becomes more aerobic off-site (increased ORP)
 - Separate source?
 - Mobility of PFOA > PFOS; example of fractionation?
 - Bio-oxidative transformation?

PFOA Relative to PFOS and PFHxS and ORP with Respect Distance from Former Building (Source)

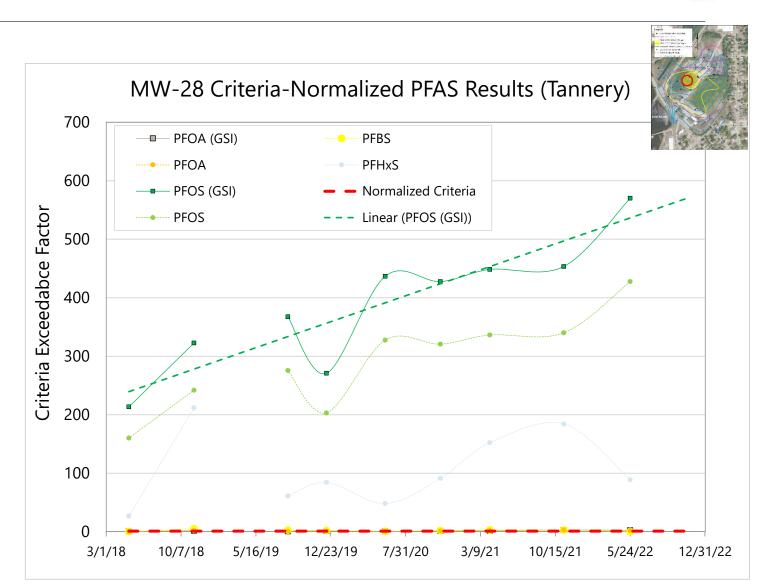


Distance from Former Building (feet)

Precursor Compounds – PFOS

The gift that keeps on giving - precursor transformation...

- 8:2 & 6:2 Fluorotelomer Sulfonate (FTS) & Et-FOSA in soil, groundwater, storm water surface water and/or foam
- PFOS concentrations are increasing in groundwater in untreated source area.
 Potential bio-transformation?



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Biodesktop 1.0 Biodesktop 2.0

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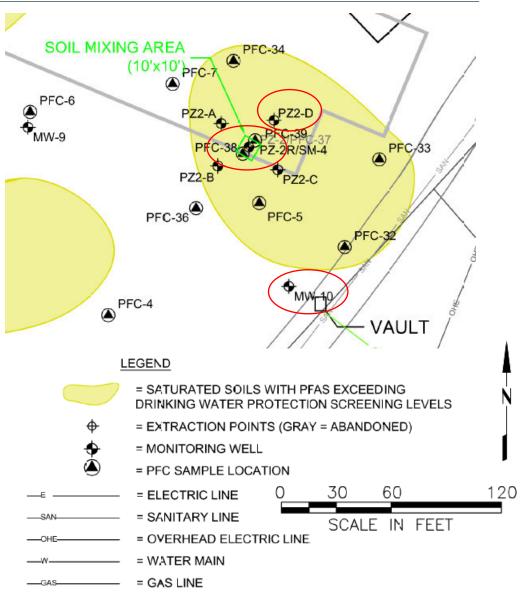
Microbial Benchtop Study – Seeking Endemic Opportunists

Why Microbes?

- An in-situ destructive mechanism is needed
- Microbes are resilient and adaptable
- The hunt for microbes is never easy (e.g., TCE, 1,4-Dioxane)
- Evidence building for bio-defluorination of PFOS and PFOA (e.g., Huang and Jaffe, 2019; Harding, 2023?)

2020 - Collected soil and groundwater to identify potential endemic, PFAS-degrading microbes (Fixed Earth LLC):

- Upgradient of soil mixing area (PZ-2D)
- Soil mixing area (PZ-2R)
- Aerobic transition zone where PFOA
 begins to increase (MW-10)

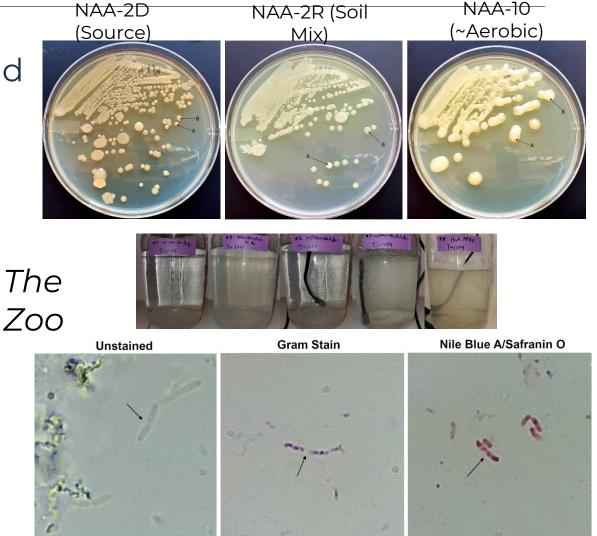


BioDesktop 1.0

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- Fixed Earth isolated 6-endemic microbial candidates from Site soil and groundwater
- Mixed microbial strains in Site groundwater:
 - Control (C);
 - Microbes (only; M);
 - Microbes + sugar + aeration (MFA)
- Mixed microbial strains in spiked PFOS/A tap water (1,000 ng/L ea.)
 - Control (C)
 - Microbes (only; M)
 - Microbes + sugar (MF)
 - Microbes + aeration (MA)

Microbes + sugar + aeration (MFA)



Production of polyhydroxalkanoaes (PHA) suspected to give apparent positive gram g test

BioDesktop 1.0 – Tap Water Results



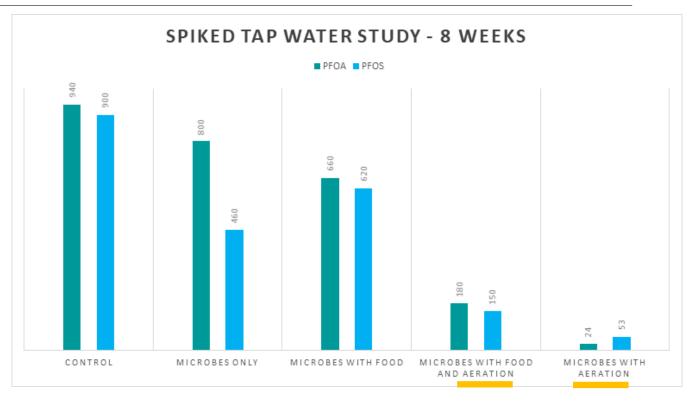
 PFOA/PFOS show significant reduction compared to control at 8 weeks:

94% to 97% PFOS/A reduction

- No fluorinated VOCs detected (open scan)
- Positive inorganic fluorine response in PFOS/A enriched media in 2 of the 6 microbes isolates.

Note yellow glow in "biofilm"?

 No significant change in inorganic water chemistry

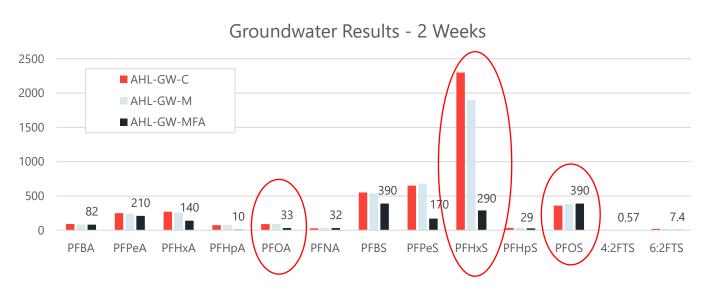




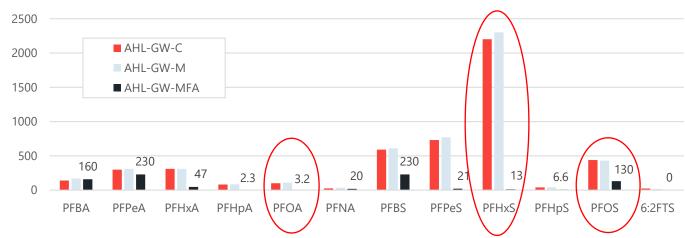
BioDesktop 1.0 – Groundwater Results

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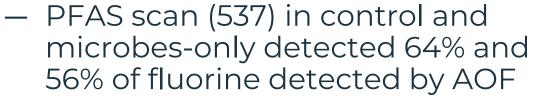
- Significant drop in PFHxS at 2- and 8-weeks with aeration
- At 2-weeks PFOS increased in MFA reactor
- At 8-weeks PFOS decreased in MFA
- Microbial augmentation alone showed modest changes – more reduction observed with aeration
- PFBA and PFPeA were only compounds to increase in MFA reactor at 8-weeks



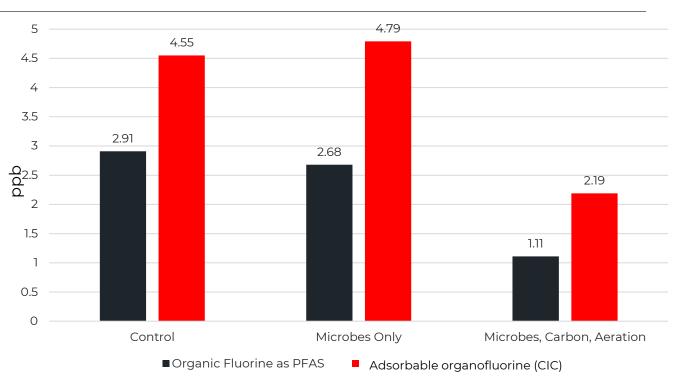
Groundwater Results - 8 Weeks



BioDesktop 1.0 – Groundwater "Precursor" Results



- PFAS scan (537) in MFA detected
 51% of fluorine detected by AOF
- 42% of the AOF reduced in the MFA sample when compared to control & microbes only
- AOF is "non-unique" but potential precursor transformation loss
- AOF does not reflect fluorine from short chains (<C4; sorption process)
- Precursor transformation may have contributed to PFOS increase at 2-weeks?

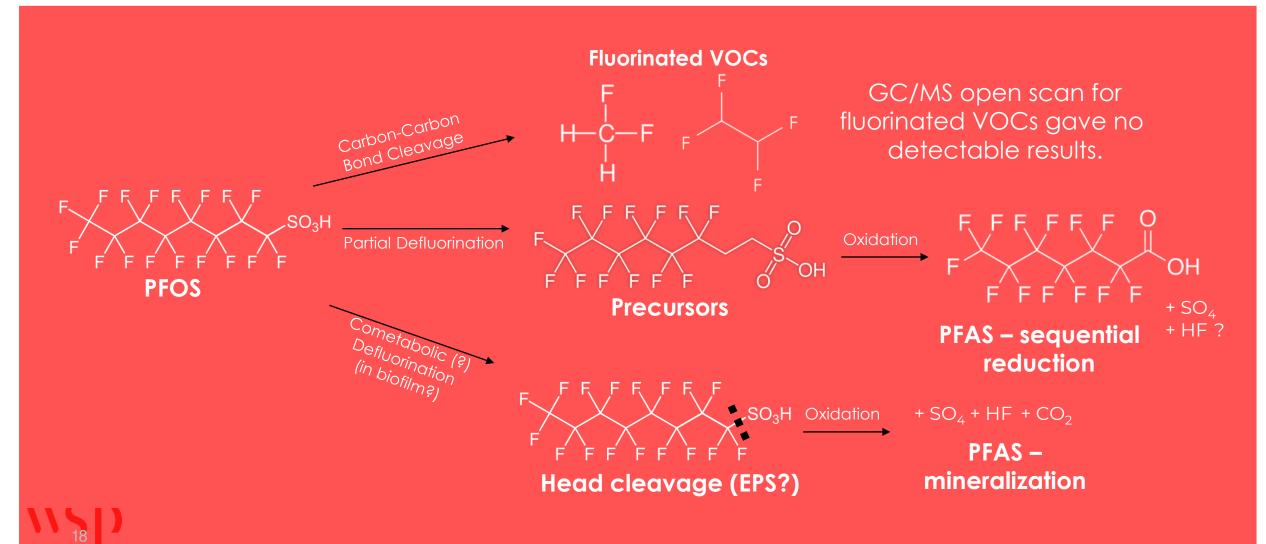




BioDesktop 1.0 – Roadmaps to Destruction?

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Mineralization model best fits observed results



PFOS/A Microbial Degradation Isn't Supposed to Happen

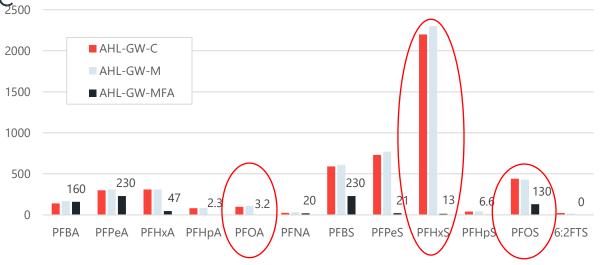


Conventional wisdom: Aerobic biodegradation is "unlikely" -what "went wrong"?

Q. Could aeration have caused loss via aerosol production or partitioning?

A. Potentially – we did not have a control on aeration only

- Q. Used Method 537 (not whole bottle) extraction techniques container loss?
 - A. Controls which should also have been affected but were not and this does not explain the breadth of data.
- Q. Work was not completed in triplicate A. Fair point (but within budget ^(C))
- Q. No "killed" control to assess bioadsorption?
 - A. Differences in multiple microbe runs suggest it was not a factor



Groundwater Results - 8 Weeks

WSP and Fixed Earth Sponsored

- Freezer stock of isolated microbial strains cultured and added to PFOS/A spiked tap water.
- Single (1-week) & duplicate (2-week) analyses
- 537 Method with whole bottle extraction
- Viability & qualitative inorganic fluorine testing
- Dose 1.25 Million CFU/mL
- Additional controls added (including heat killed microbes [HKM] to assess bio-adsorption)
- Rock flour (RF) added to assess role of a substrate
- Use of a diffuse oxygen source, shaker table & peroxide (partitioning/aerosol concerns)









BioDesktop 2.0

BioDesktop 2.0/2.1 – Results at 2-weeks



- Microbes in aerated samples were viable
- Viable colonies then demonstrated positive inorganic fluorine response
- HKM (bio-adsorption: 0-12% Reduction)
- Partitioning (14-32 % diff between top & bottom)
- Live microbe reductions of 0-12%







Viability test (growth); PFASenriched media, 6 E08 CFU/mL 2.0: PFOS 1200 ng/L; PFOA 1400 ng/L

2.1: PFOS 3000 ng/L; PFOA 800 ng/L

2.0: 2-Week results; diffusive oxygen source

	Average Duplicate Results at 2-wks (ng/L)					
Condition	Control		Live Microbes			
	PFOS	PFOA	PFOS	PFOA		
Stock Solution	1000	1300				
Rock Flour (RF)	1100	1400	995	1300		
Diffusive Oxygen (O2)			980	1300		
RF+O2	1100	1400	1000	1300		
RF+O2+HKM	1200	1400				
RF+Peroxide			995	1300		

2.1: 2-Week results; shaker oxygen source

	Results at 2-wks (ng/L)					
Condition	Con	trol	Live Microbes			
	PFOS	PFOA	PFOS	PFOA		
Stock (top)	3400	700	2200	660		
Stock (bottom)	2300	600				
RF+Shaker O2	3000	750	3400	830		
Shaker O2	3000	810	2900	810		
HKM (Sealed/shaken)	2500	770				

PFOS/A Microbial Degradation Was Supposed to Happen

Bio-desktop 2.0 Wisdom: Aerobic biodegradation "worked"-what "went wrong"?

- Q. Could aeration have caused loss via aerosol production/partitioning in 1.0?
 - A. Potentially we did not have a control on aeration only during this experiment (**but will in 3.0**)
- Q. Container loss?
- A. 1.0 controls still should have been affected.
- Q. "Killed" control to assess bio-adsorption?
 - A. Killed microbe sets suggest 0-12% adsorption (including standard 537 extraction to isolate biofilms from aqueous media). Biofilm consideration.
- Q. What was different?
 - A. Refined microbial strain isolates used in 2.0; was something "lost" that caused enzyme expression? Fresh samples to be used for Bio-desktop 3.0

Dr. Chris Marshall (Marquette University, WI) performing third party, independent validation & genome mapping

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This is driving me crazy – let's go outside 2021 & 2022 Pilot Testing

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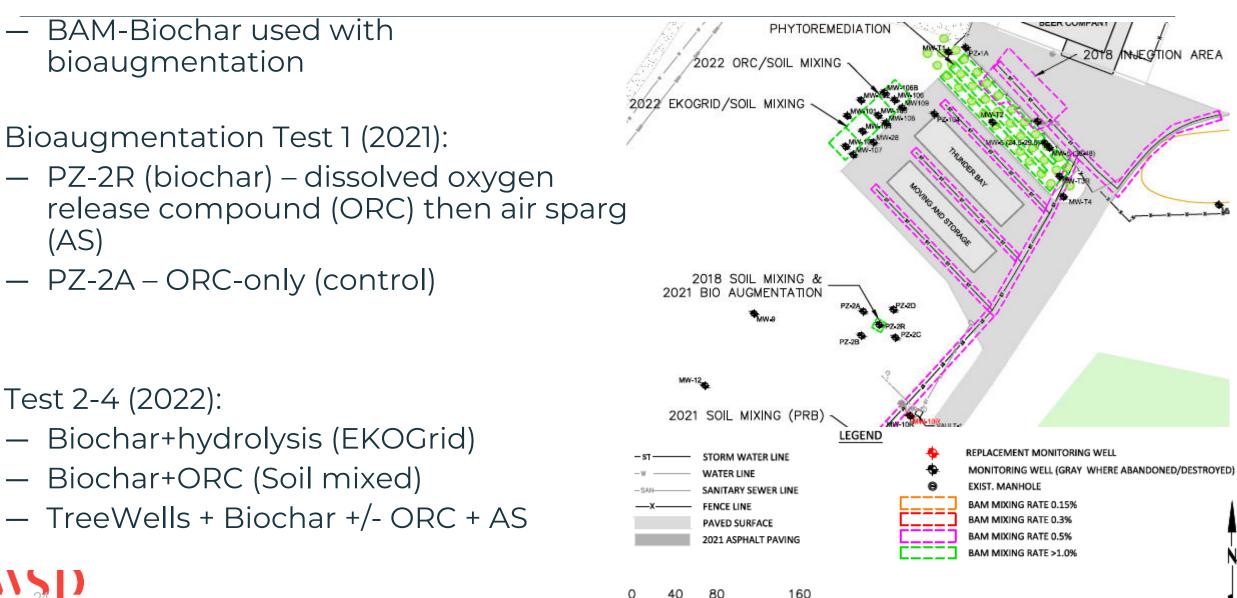
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Microbial Pilot Tests

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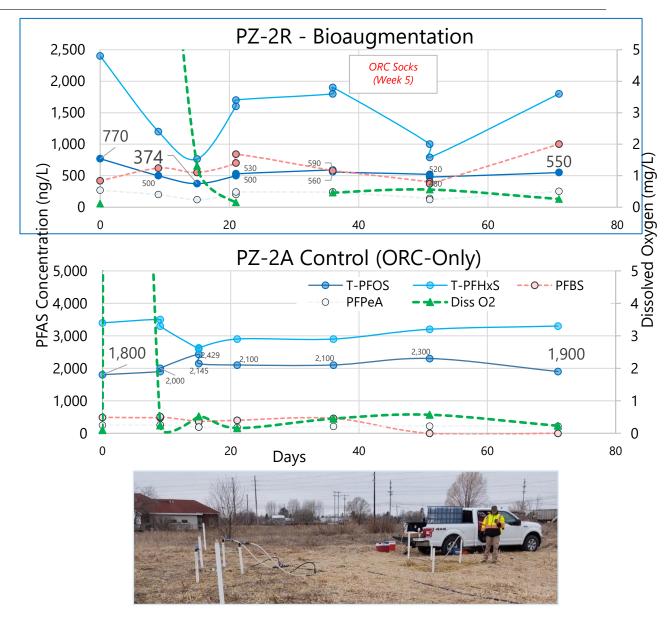


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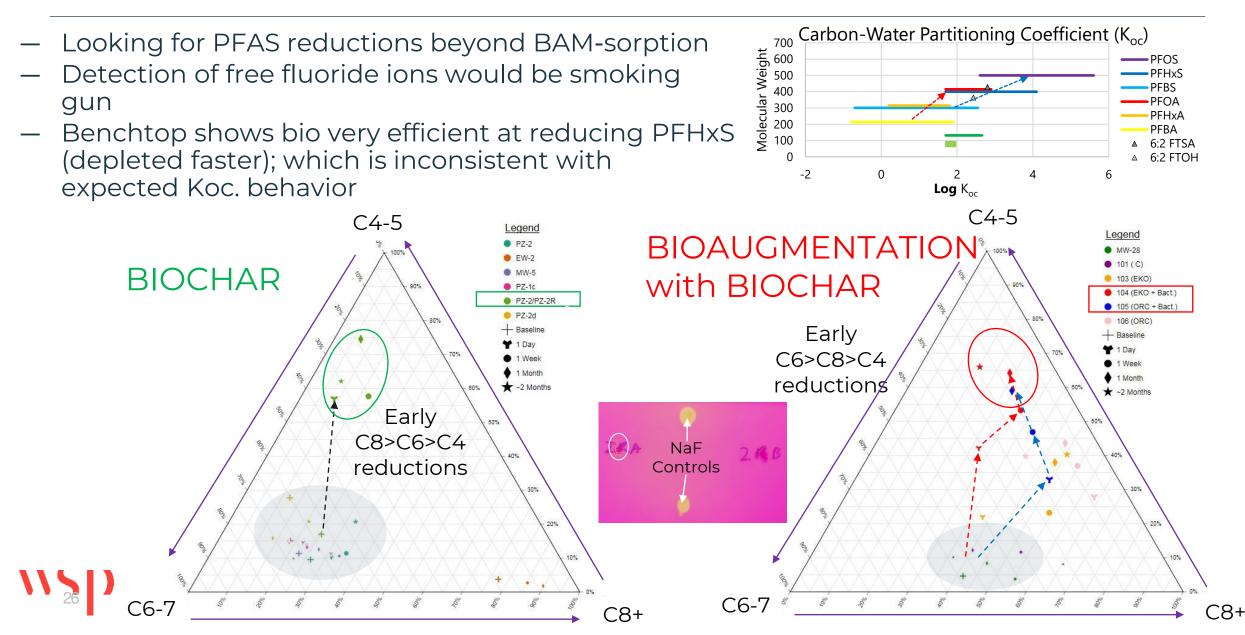
Microbial Pilot Test – 2021 Results

- Delivery via three dedicated injection wells in treatment (PZ-2R) and control (PZ-2A) blocks
- Bioaugmentation: Significant declines in PFHxS & PFOS in first 2weeks
- PFAS rebound when DO << 1 mg/L (AS added later)
- Short-term reduction inconclusive but declines > than ~10% bioabsorption observed in HKA (Biodesktop 2.0)
- PFOS increased in ORC-only control Precursor "source" of PFOS masking breakdown?
- Displacement unlikely cause of
 beserved reductions



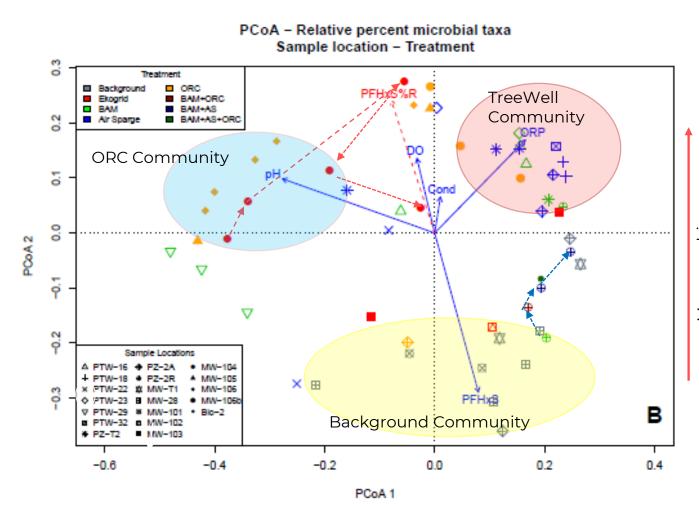
Is there a "Bio" signature?





Is there a better way to assess aquifer buffering?

- Principal component analysis to statistically assess community similarities and differences
- Allows environmental parameters (Eigen vectors) to map across PCoA field to assess role of aquifer conditions on microbial communities.
- Can also be used to assess PFAS concentration (or percent reductions)
- Microbial communities change in response to treatment approaches



More aerobic

Summary – Biochar / Bio-desktop results

- Probably not a single microbe "silver bullet"
- Biodegradation results are inconclusive (1 for 2):
 - Microcosm Benchtop 1.0 provided compelling data suggesting degradation (PFAS mineralization and liberation of free fluoride) may occur under enhanced aerobic conditions.
 - Microcosm Benchtop 2.0 failed to show significant biodegradation or bio-adsorption processes.
 - Air-water partitioning NOT significant to explain lack of reduction
 - Processed strains lost key element for necessary gene expression?
 - Desktop 3.0 will use site groundwater and provide additional control on aeration mechanism
 - 3rd party validation & genome mapping underway
- Microbes unable to be "separated" during genome mapping

- Mass balance continues to pose challenges and requires reliance on a multiple lines of evidence approach.
 - AOF and fluorinated VOC open scans suggest that large chain precursors were reduced and no cleavage of PFAS (C-C) evident (Desktop 1.0)
 - Better quantification of inorganic fluoride is one of the elements needed to improve mass balance control.
 - Qualitative results suggest microbial enhanced mineralization of PFAS occurred (Desktop 1.0 and in viable samples at 2-weeks in desktop 2.0/2.1).



Summary – Biochar / Bio Pilot

- 2021 Pilot mirrors Desktop 1.0: Early reduction in PFHxS linked to microbial degradation (microcosm and field results)? Partitioning onto carbon should favor PFOS over PFHxS.
- Precursor biotransformation to PFOA/PFOS may mask degradation of PFOS/PFOA (microcosm and PZ-2A field results)
- Bioaugmentation pilot results are inconclusive
 - Short term reductions in PFAS (Oxygen was limiting factor)
 - See decreased PFHxS relative to longer/shorter chains (ala benchtop)
 - Is there a critical loading rate? (e.g., Dhc 10⁴)
 - Reductions exceeded benchtop bio-adsorption ranges
- Verifying biologic destruction of PFAS in the field remains extremely challenging.



 Current state of the knowledge and technology is that microbialrelated remediation is not there. But the NEED is there. These studies don't disapprove or prove that, but it hopefully helps to get the ball rolling.



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Thank you!

