

Applications of Anaerobic Petroleum Hydrocarbon Bioremediation



Jeff Roberts², Courtney R. A. Toth¹, Shen Guo¹, Nancy Bawa¹, Sandra Dworatzek², Jennifer Webb², Rachel Peters³, Kris Bradshaw³, Elizabeth A. Edwards¹, Krista Stevenson⁴, Colette McGarvey⁴ and Ada Wang⁴

¹Department of Chemical Engineering & Applied Chemistry, University of Toronto, Toronto ON, Canada

²SiREM, Guelph ON, Canada

³Federated Co-operatives Limited, Saskatoon SK, Canada

⁴Imperial Oil

Acknowledgements – Benzene/GAPP Team

Dr. Elizabeth Edwards, Dr. Courtney Toth, Shen Guo, Nancy Bawa, Charlie Chen, Johnny Xiao, Dr. Olivia Molenda, Elisse Magnuson, Chris Shyi, and Kan Wu
Chemical Engineering and Applied Chemistry, University of Toronto

Sandra Dworatzek, and Jennifer Webb
SiREM, Guelph ON

Dr. Ania Ulrich, Korris Lee, and Amy-Lynne Balaberda
Civil and Environmental Engineering, University of Alberta

Dr. Neil Thomson, Andrea Marrocco, Griselda Diaz de Leon, Bill McLaren, and Adam Schneider
Civil and Environmental Engineering, University of Waterloo

Dr. Karen Budwill, and Stanley Poon
Innotech Alberta, Edmonton ON

Krista Stevenson
Imperial Oil Limited, Sarnia ON

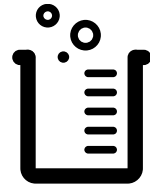
Kris Bradshaw, and Rachel Peters
Federated Co-Operatives Limited, Saskatoon SK



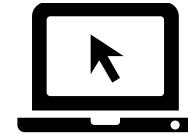
Introduction to SiREM



Founded in 2002 in
Guelph, ON



Provide products
and testing services
to support and improve site
remediation



Further information:
siremlab.com



SiREM Service Areas

Treatability Testing



treatability
studies

SiREMNA™

Characterization/Monitoring

- *Molecular Testing*

gene & trac®



- *Passive Samplers for Vapor and Pore Water*

Bioaugmentation

KB-1®

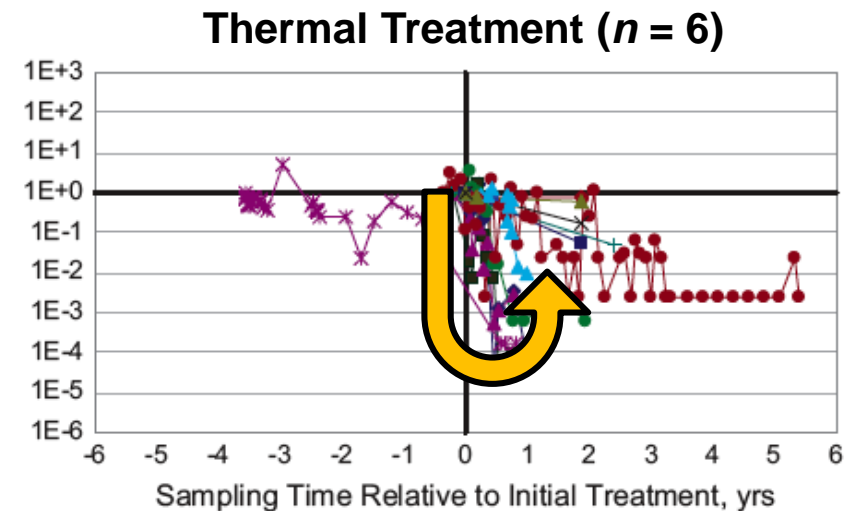
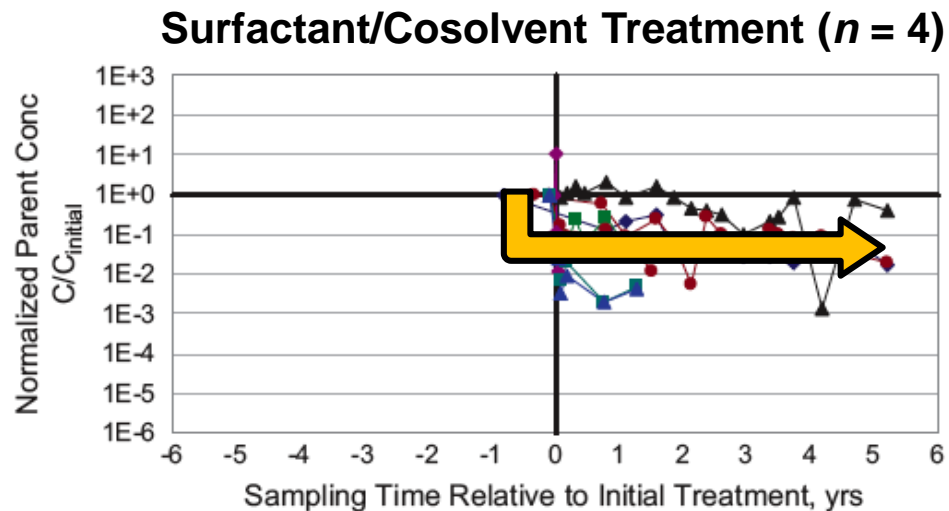
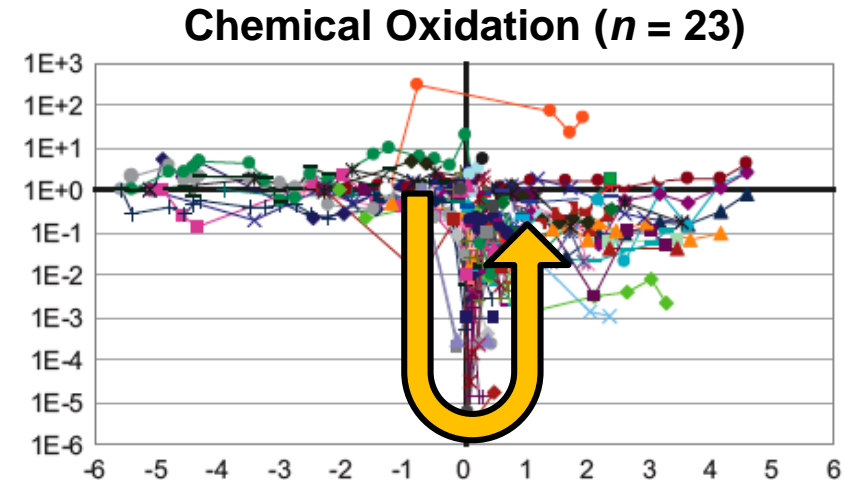
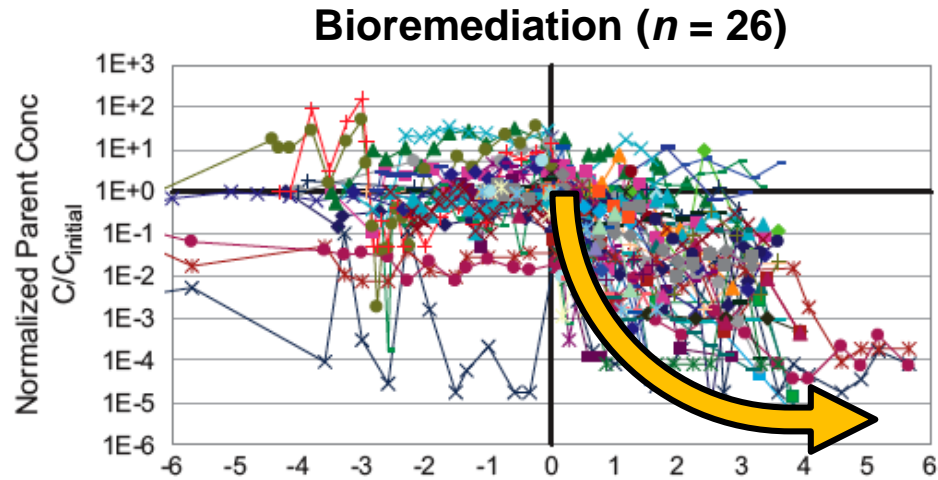
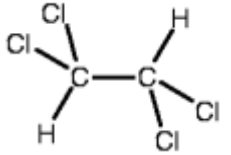
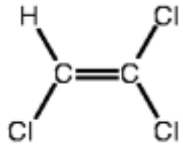


WATERLOO
MEMBRANE
SAMPLER



SP3

■ ■ Bioaugmentation has been an effective treatment for chlorinated
■ ■ solvents. Can it be used to treat other contaminants





Why Go Anaerobic for BTEX?

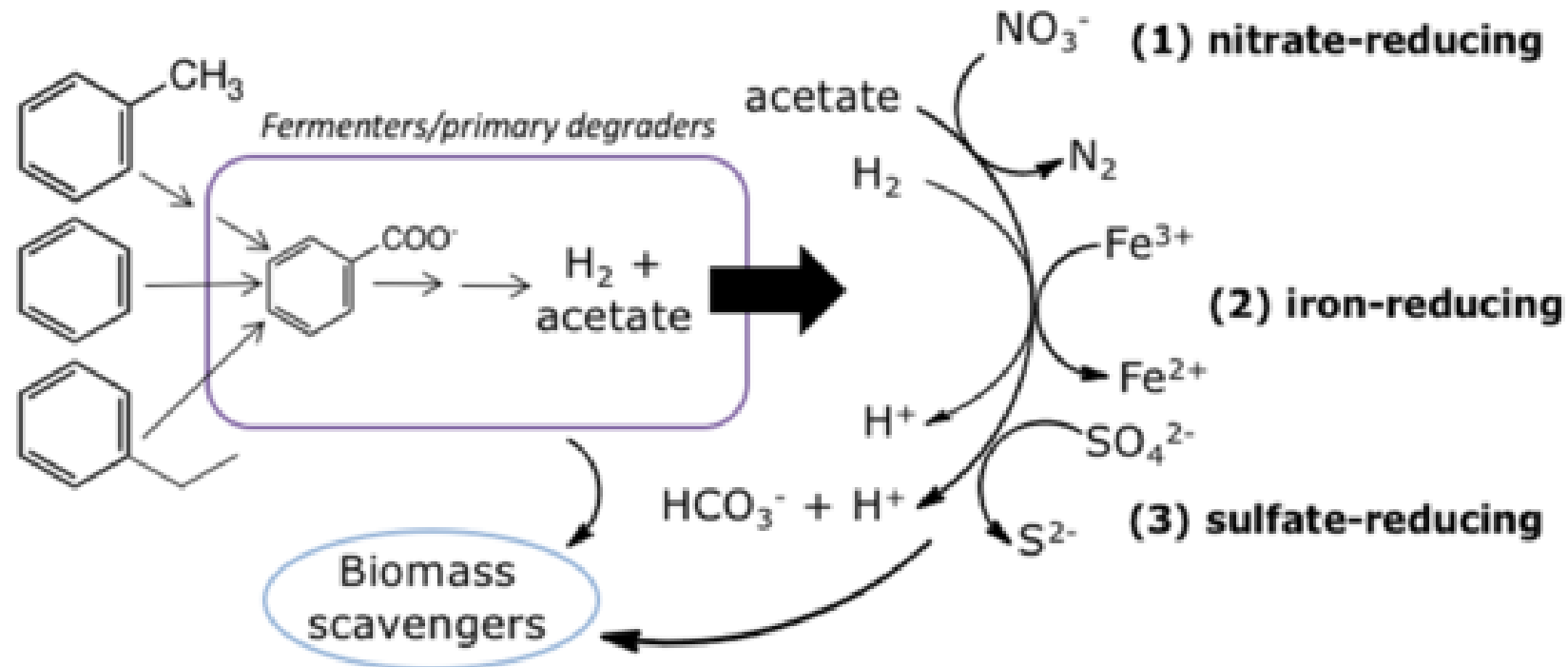
- Hydrocarbon sites can go anaerobic - high organic loading consumes O_2
- Electron acceptors ($NO_3^-/SO_4^{2-}/CO_2$) often already present in subsurface
- Anaerobic electron acceptors are soluble, easier to apply/distribute compared to O_2 (e.g., epsom salts (sulfate))
- Viable *in situ* remediation option for deep contamination



Key Difference Between Bioremediation of Chlorinated Solvents vs Hydrocarbons

Hydrocarbons are *electron donors* rather than electron acceptors

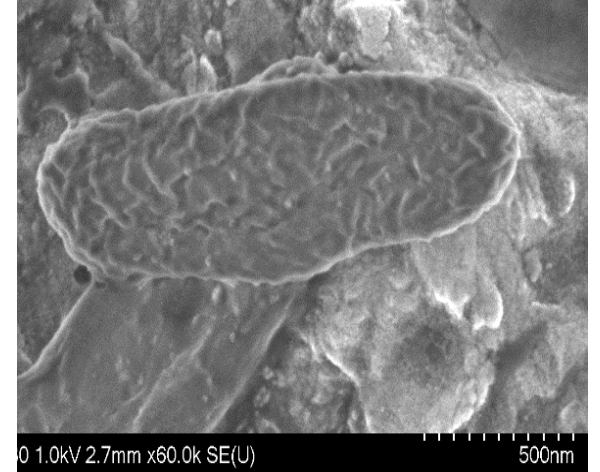
- Adding carbon (sugars, VFAs, yeast extract) may not enhance bioremediation performance
- Adding electron acceptors does not always enhance bioremediation either





ORM2 Anaerobic Benzene Degradere

- Benzene specialist derived from an oil refinery site in 2003
- ORM2 is a *Deltaproteobacterium*
- Produces enzymes that ferment benzene
- Slower growing ~ 30 day doubling time

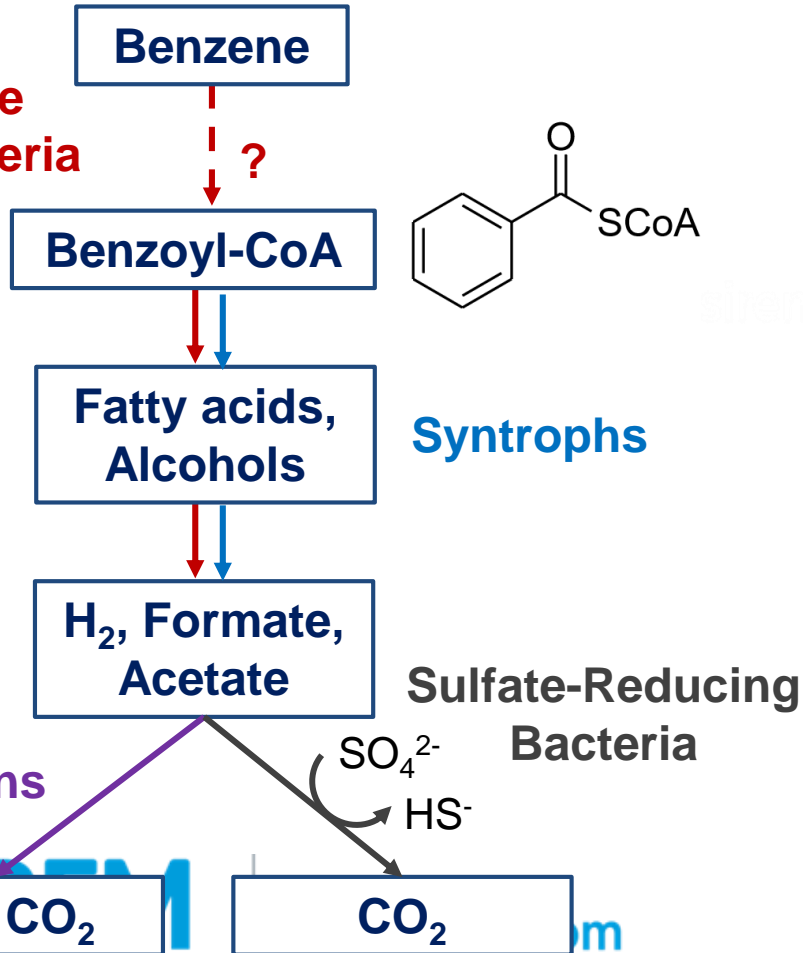




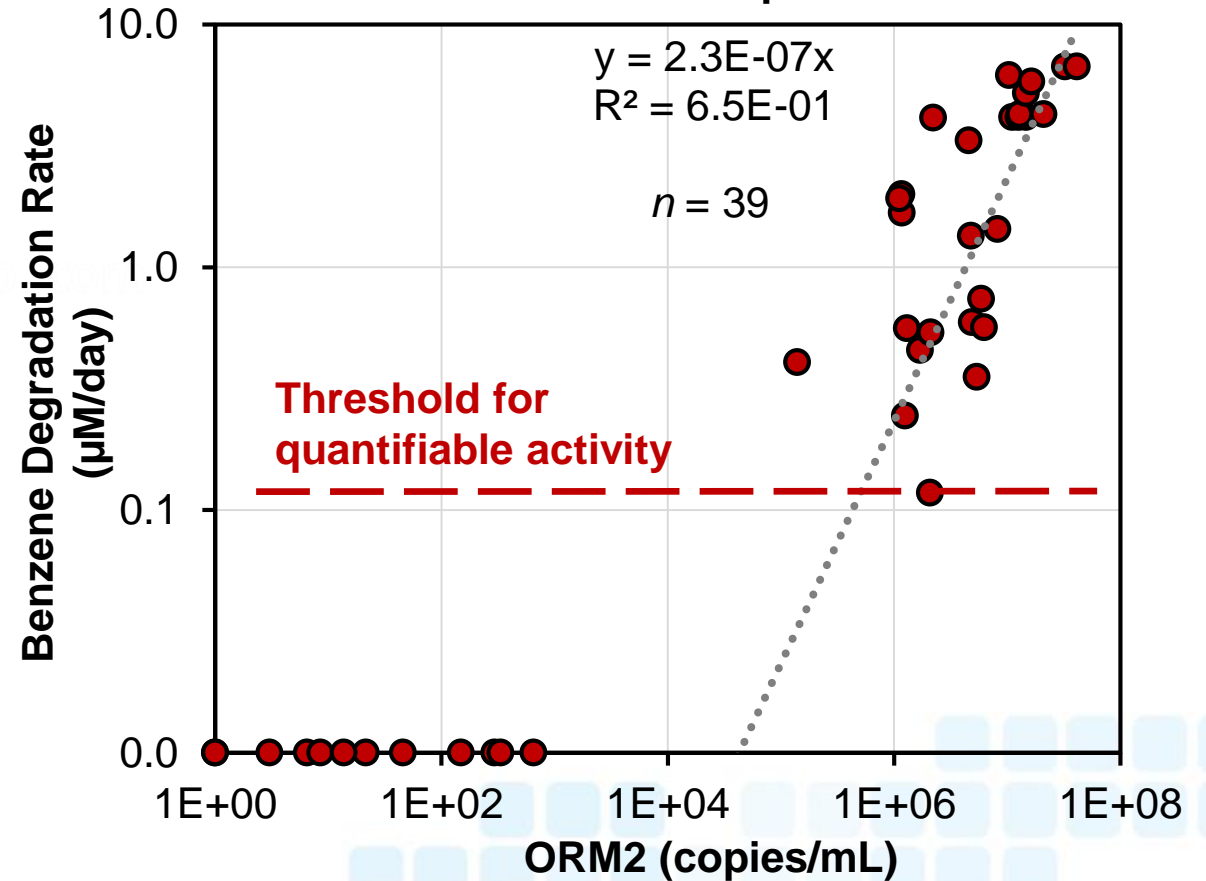
Rates of Anaerobic Benzene Degradation is Linked to Concentrations of Key Microbes



**Sva0485 Clade
Deltaproteobacteria
(ORM2)**



0.1 μM benzene/day = 7.8 μg benzene/L/day
= 4.3E+05 ORM2 copies/mL



Treatability Study Experimental Setup

BTEX contaminated

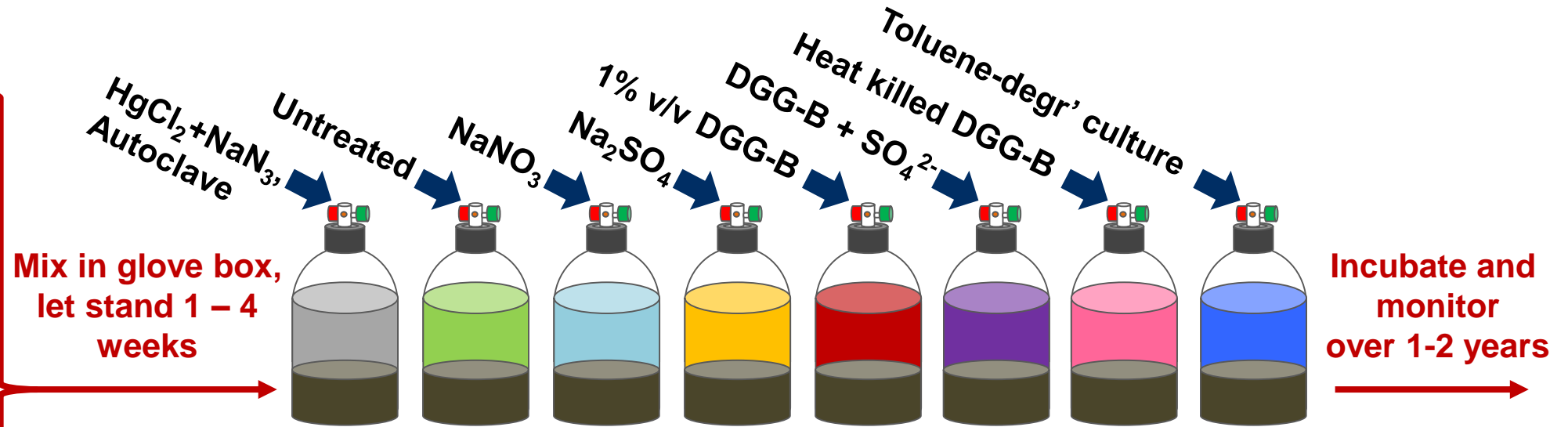


**Crushed core sample
(60 g)**



**Groundwater sample
(200 mL)**

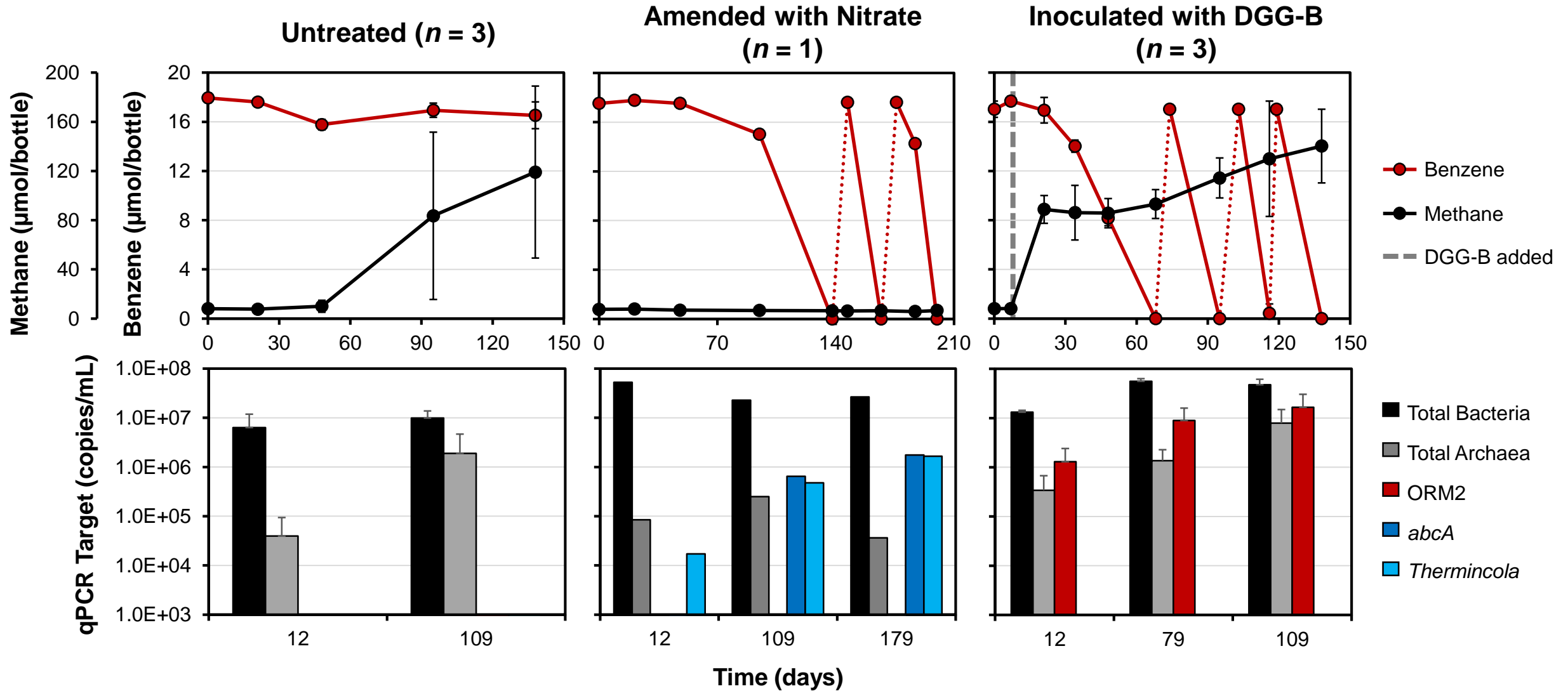
Aqueous BTEX concentrations depend on site materials



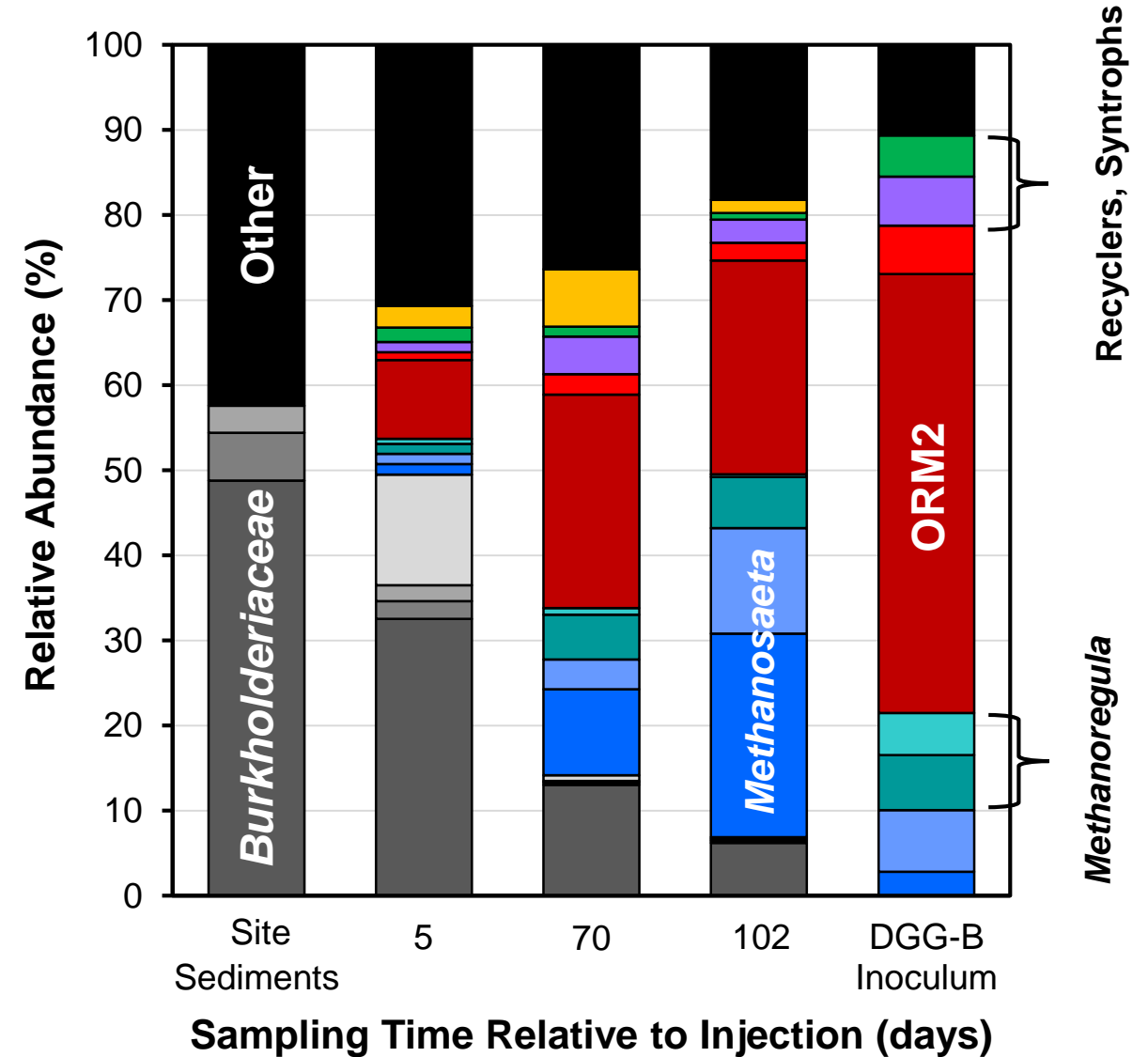
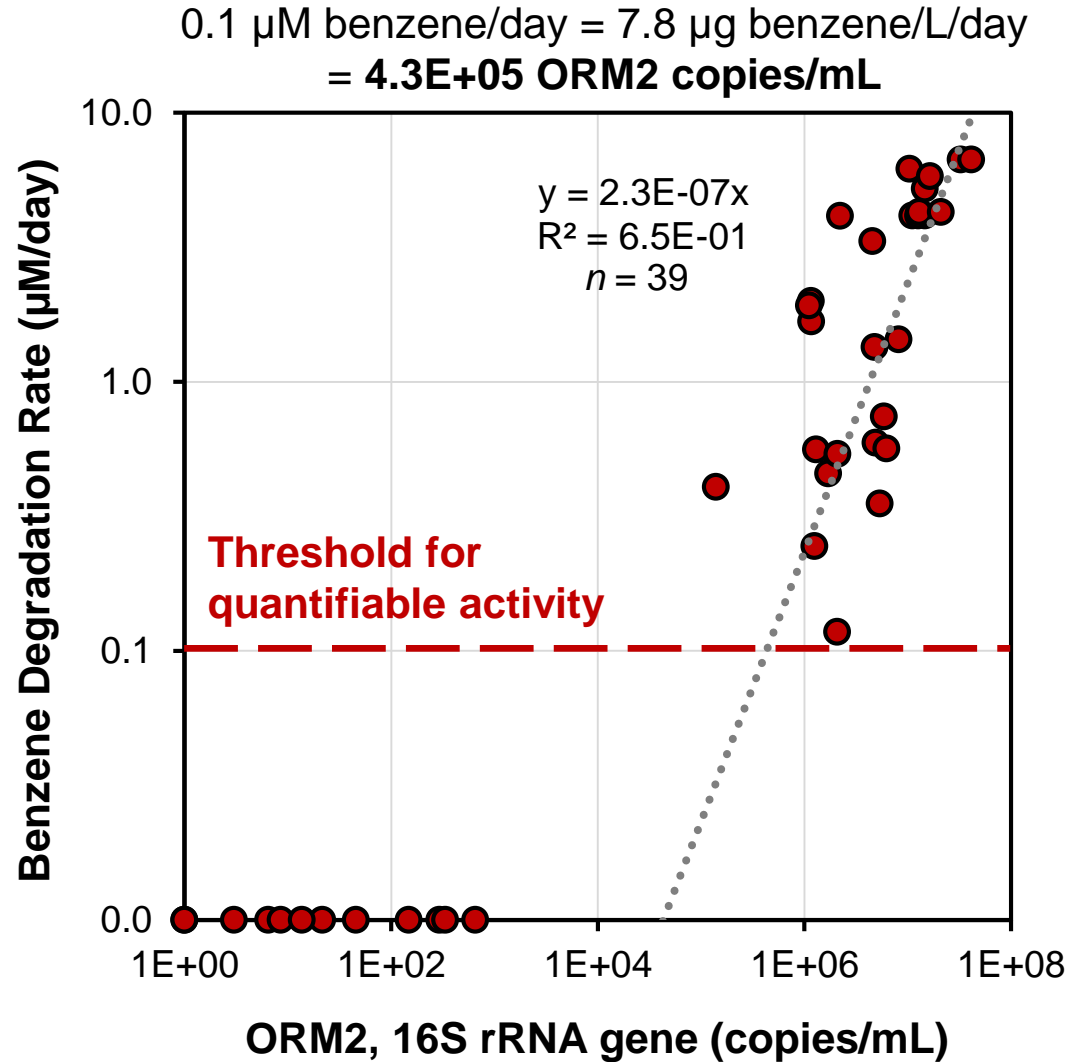
**230 mL groundwater slurries
20 mL headspace (10% CO₂ / 90% N₂)**



Treatability Test Results

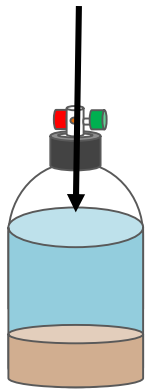


DGG-B Genomic Monitoring Tools

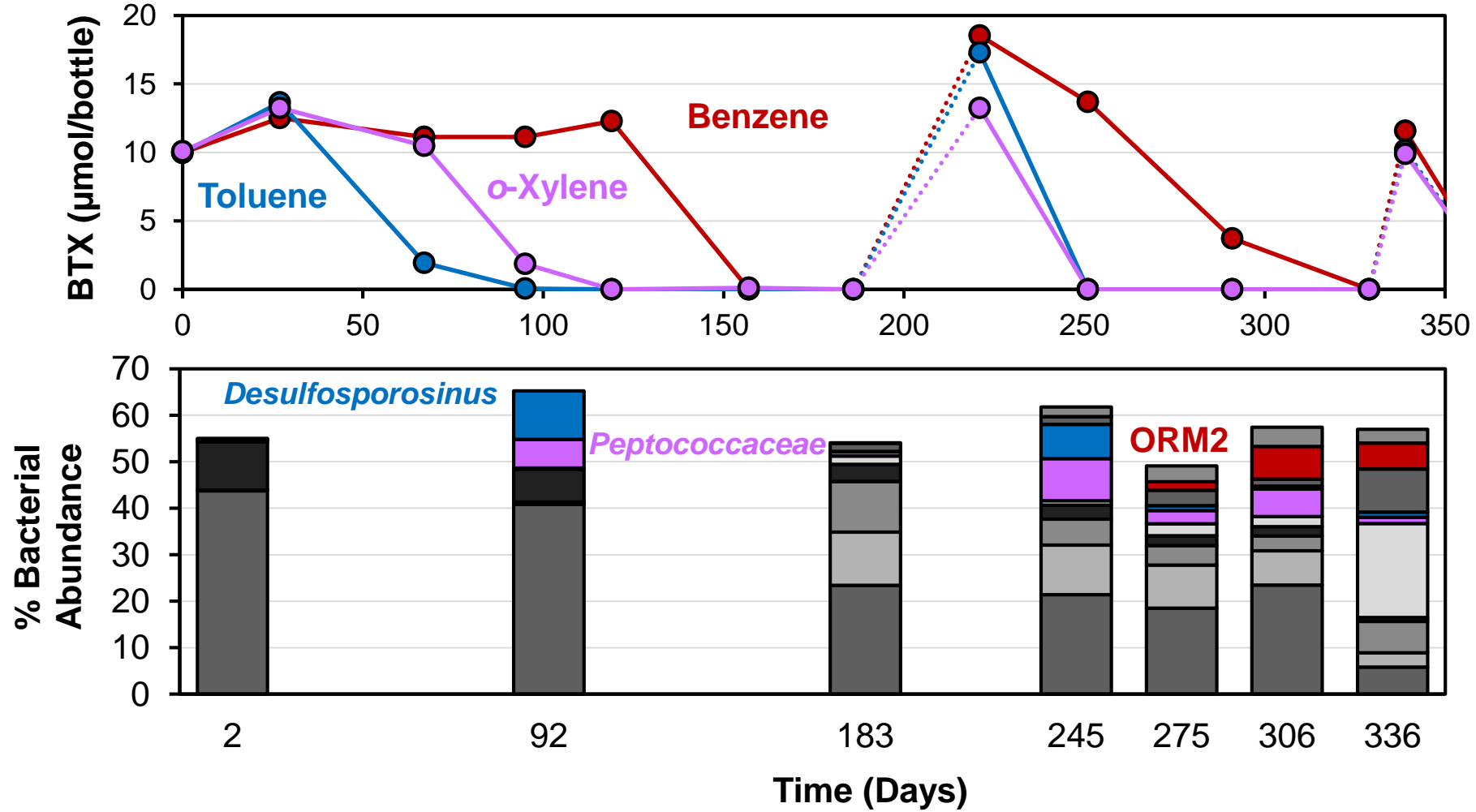


Concentrations of Key Microbes May Also Control Rates of Anaerobic TEX Biodegradation

Additional testing and qPCR analyses are underway

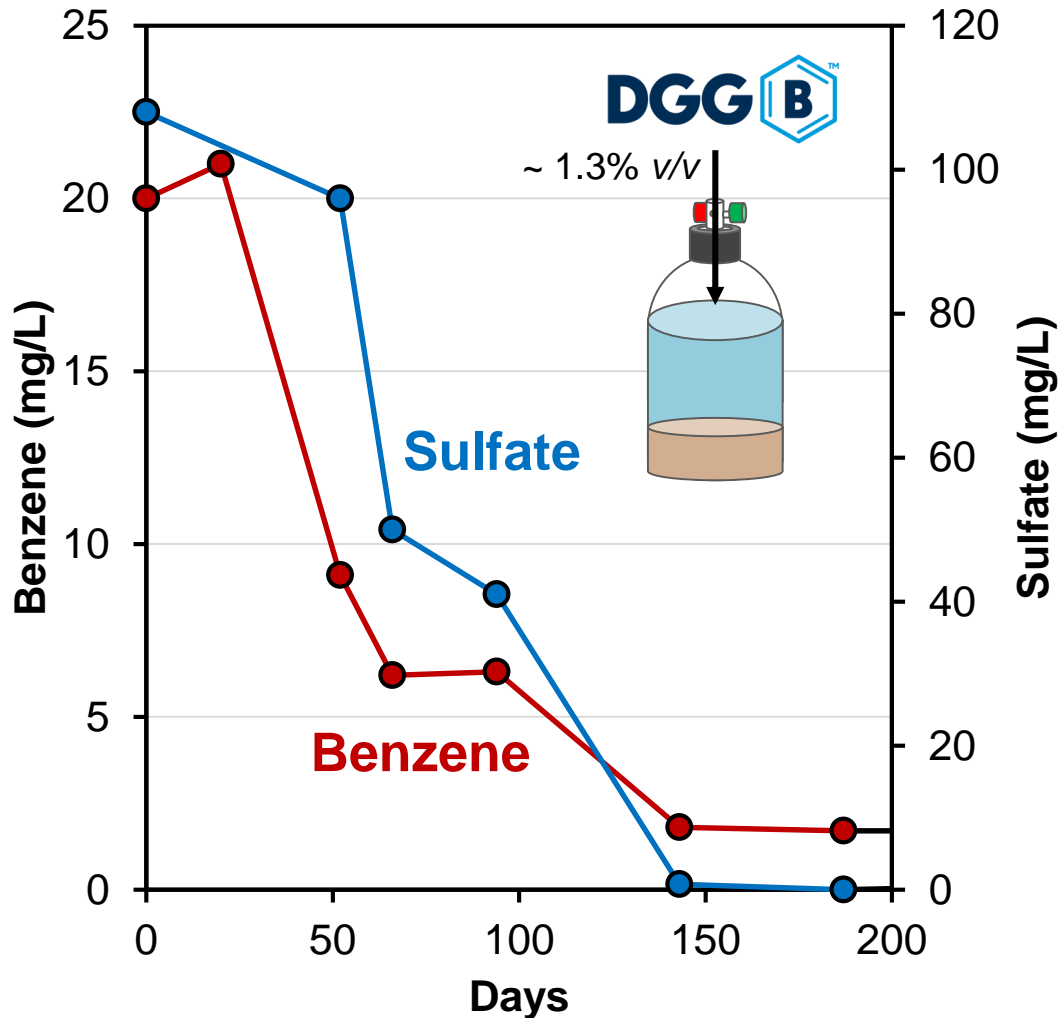
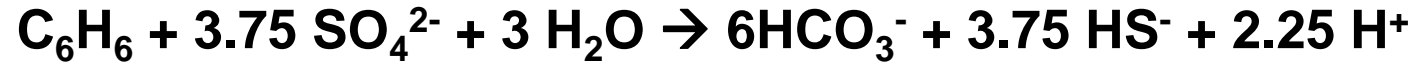


BTX-contaminated sediments and GW (~ 6 mg/L each)



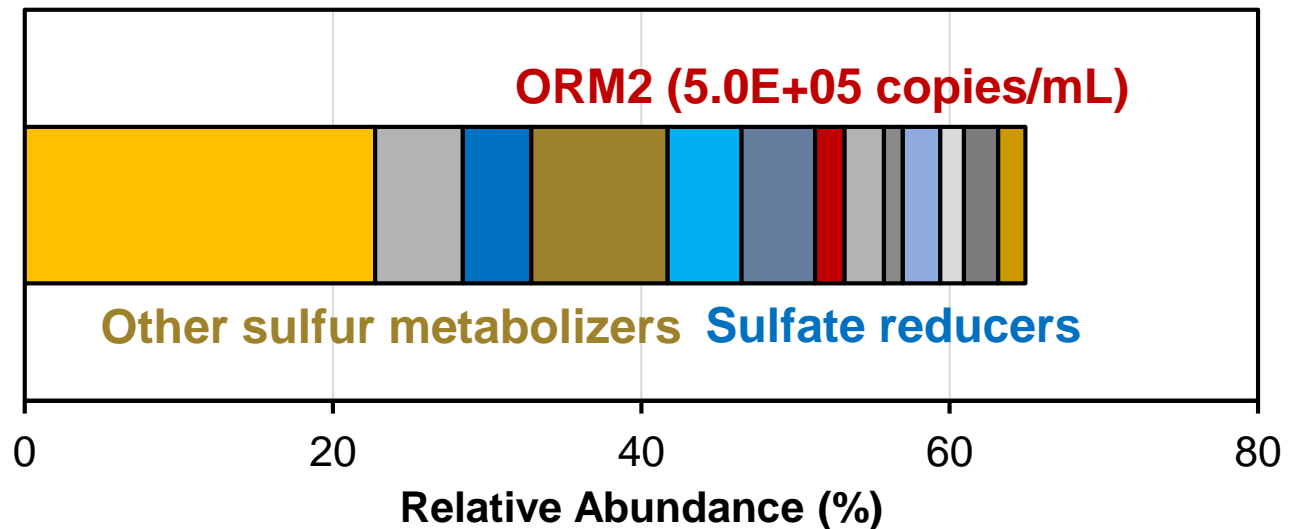
Treatability Testing with DGG-B

Benzene metabolism coupled to sulfate reduction was observed in DGG-B inoculated bottles



Total Benzene loss (μM)	Sulfate loss (μM)	Sulfate/Benzene Ratio
234	812	3.5

Microbial Community Composition after 147 days



What Limits BTEX Biodegradation in Groundwater?

Hydrocarbon Properties? ❌

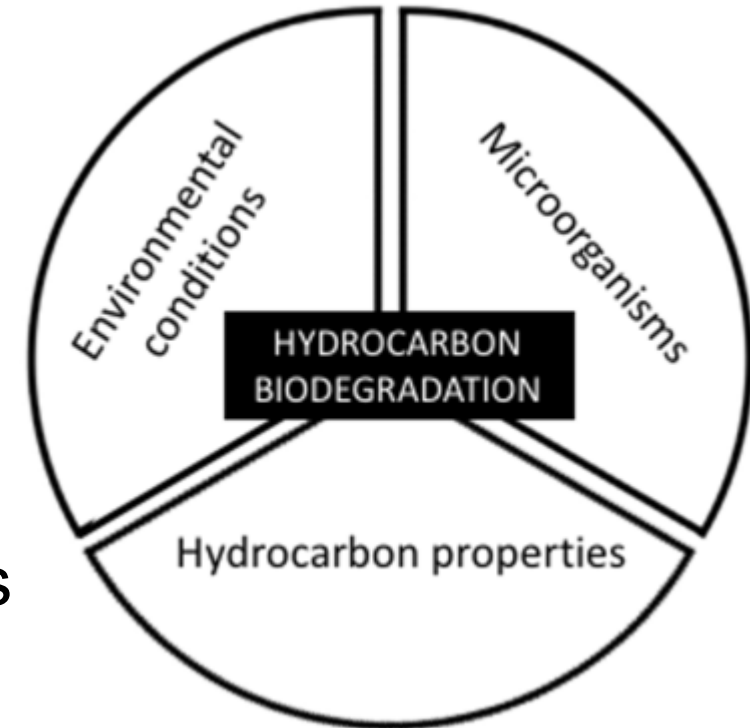
- BTEX is susceptible to biodegradation

Environmental Conditions? *unlikely*

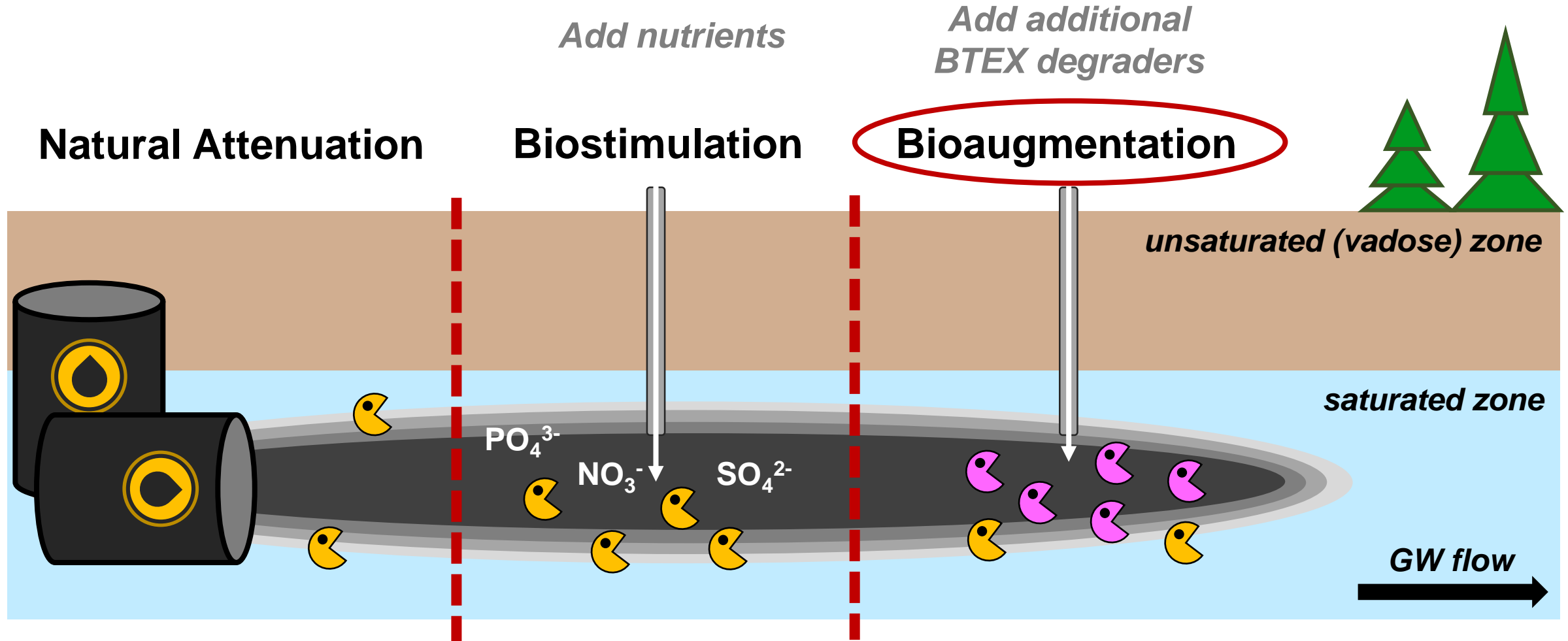
- Biodegradation occurs under all major electron-accepting conditions (O_2 , Fe^{3+} , NO_3^- , SO_4^{2-} , CO_2)
- Nutrients are recycled over time
- pH, °C, co-contaminants may ↓ degradation rates

Microorganisms? ?

- BTEX degraders are ubiquitous in nature...
- *...but they aren't always in sufficient quantities*



How Can We Reliably Increase Concentrations of BTEX Degraders?



Project Goal & Success Criteria

In field trials, demonstrate the efficacy of anaerobic bioaugmentation cultures to treat BTEX-contaminated groundwater

1. Groundwater BTEX concentrations must decrease post-bioaugmentation, relative to untreated (control) wells;
2. BTEX loss/depletion should be sustained over the posttreatment monitoring period (***years!***); and,
3. Enrichment of bioaugmented organisms (ORM2, etc.) should be evident over time.

Field Pilot Site Overview

Decommissioned gas station with historical BTEX, F1 and F2 alkane contamination

Site Overview (1993 – 2008, approximate)

Site Timeline

1993: Leaks detected from UST, oil storage, pump islands

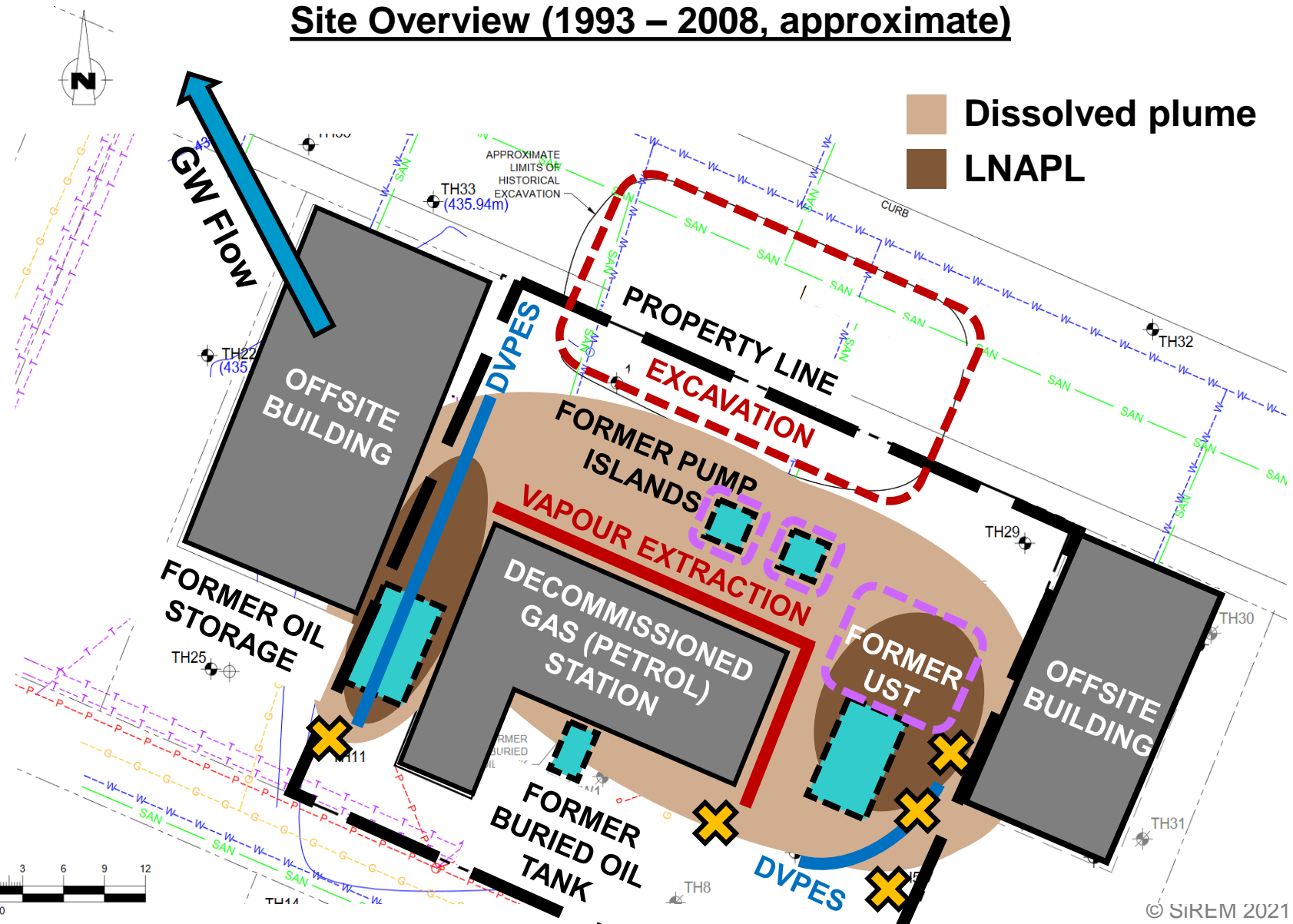
1993: Excavation, vapour extraction line installation

2005: Fertilizer injection

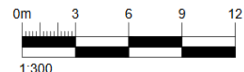
2005-'06: Dual phase vacuum extraction system (DVPES) use

2007-'08: More excavations, purging

2008: Site remediated?



Dissolved plume
 LNAPL



LNAPL = light non-aqueous phase liquid
 UST = underground storage tank

Field Pilot Site Overview

Decommissioned gas station with historical BTEX, F1 and F2 alkane contamination

Site Overview (2016, approximate)

Site Timeline

1993: Leaks detected from UST, oil storage

1993: Excavation, vapour extraction line installation

2005: Fertilizer injection

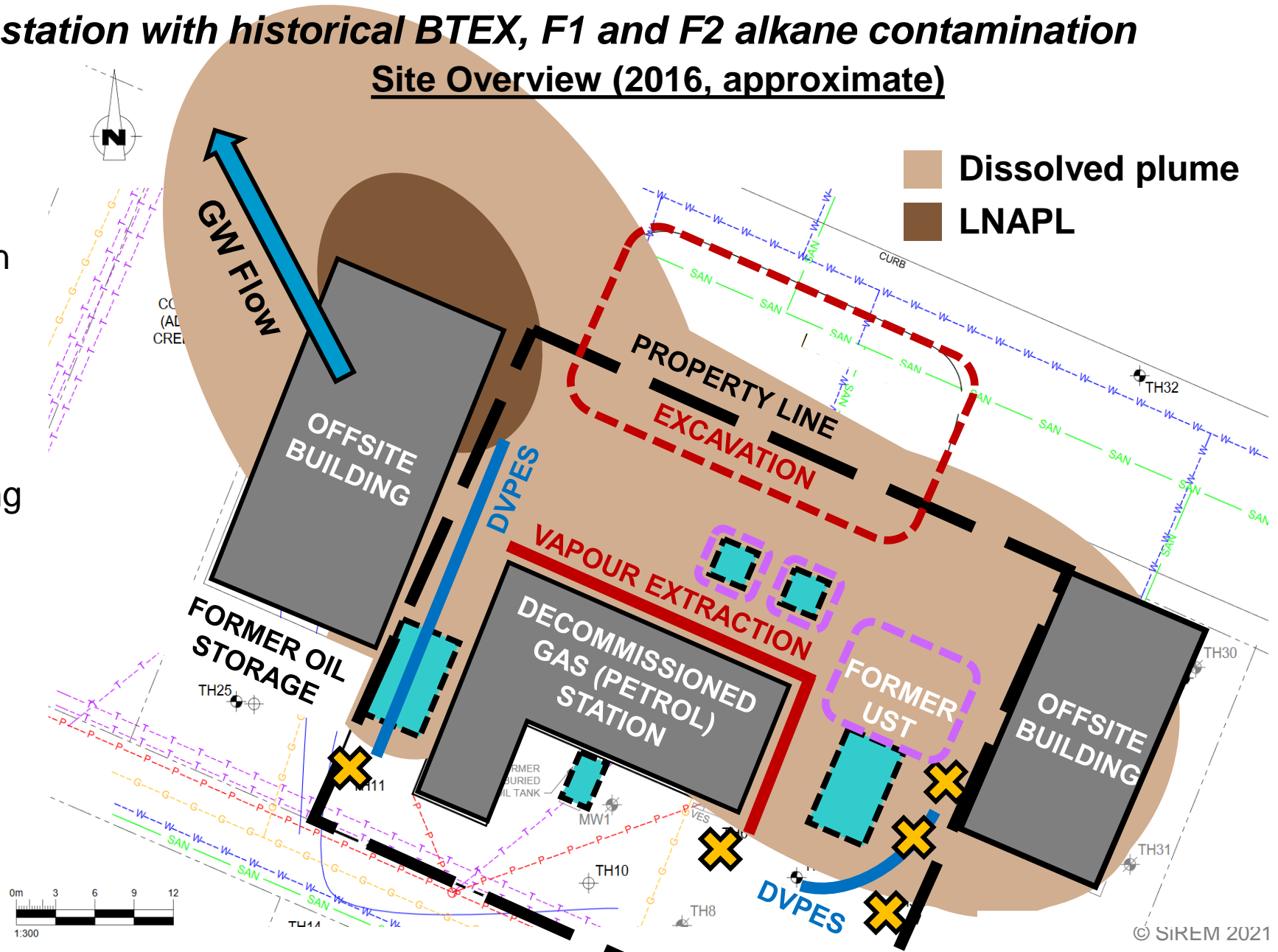
2005-'06: Dual phase vacuum extraction system (DVPES) use

2007-'08: More excavations, purging

2008: Site remediated?



LNAPL = light non-aqueous phase liquid
UST = underground storage tank



Field Injection Overview

DGG-B was injected at two direct push points (10 L each) in the LNAPL zone 5 m apart

The study was designed to treat 20,000 L of groundwater (~ 1200 ft³; 34 m³)

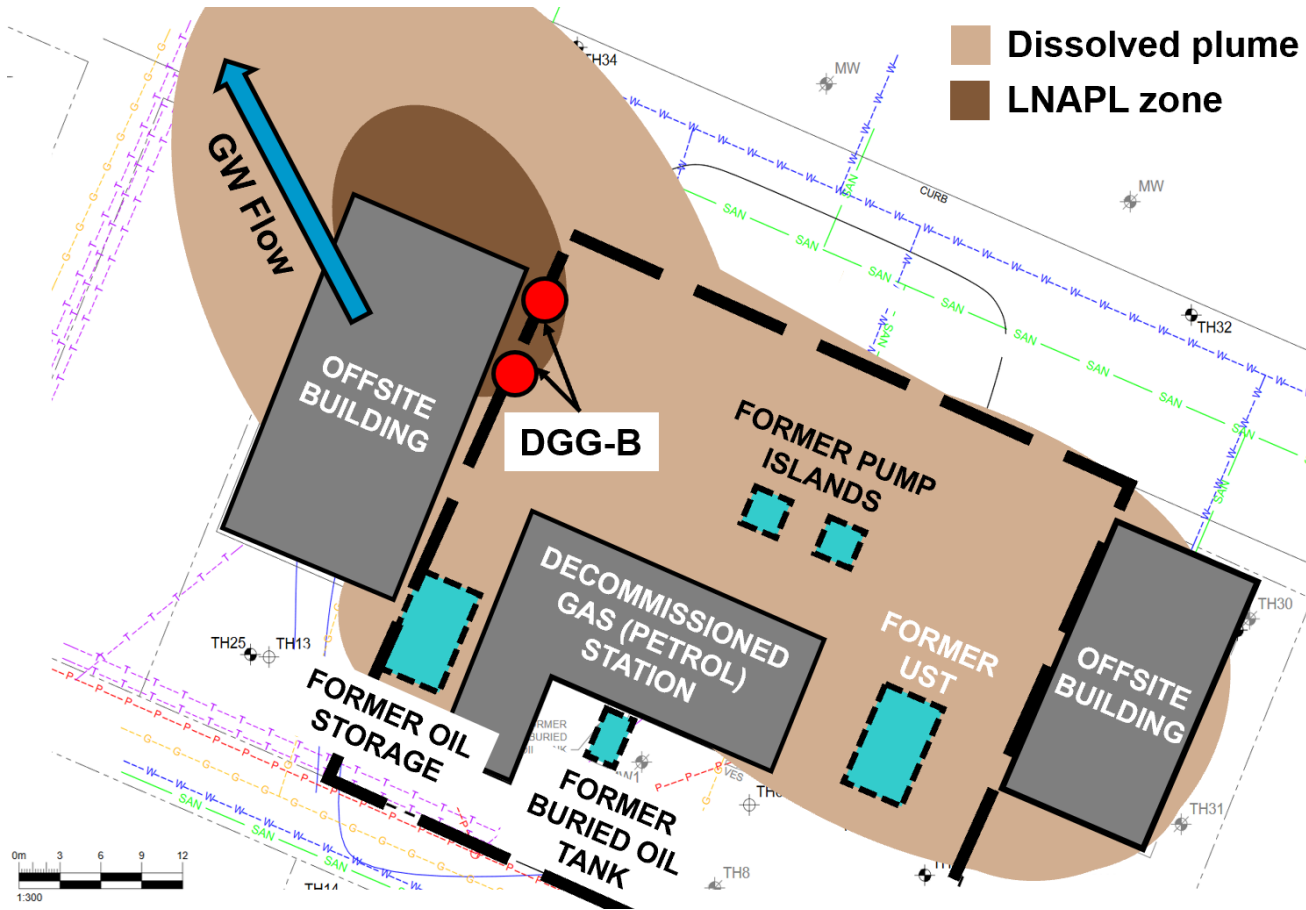
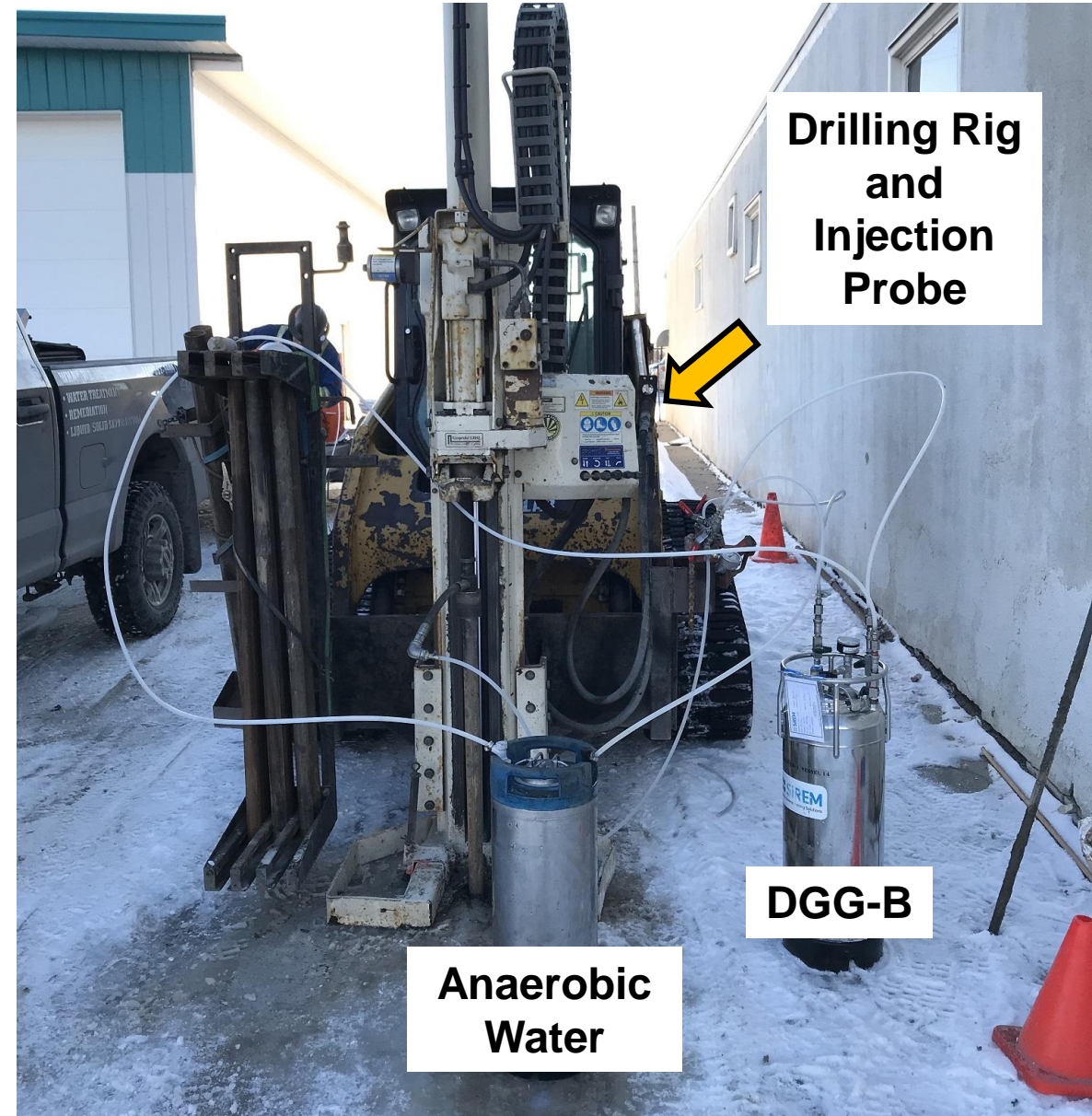


Photo of DGG-B Injection
November 14th, 2019 (-2°C)



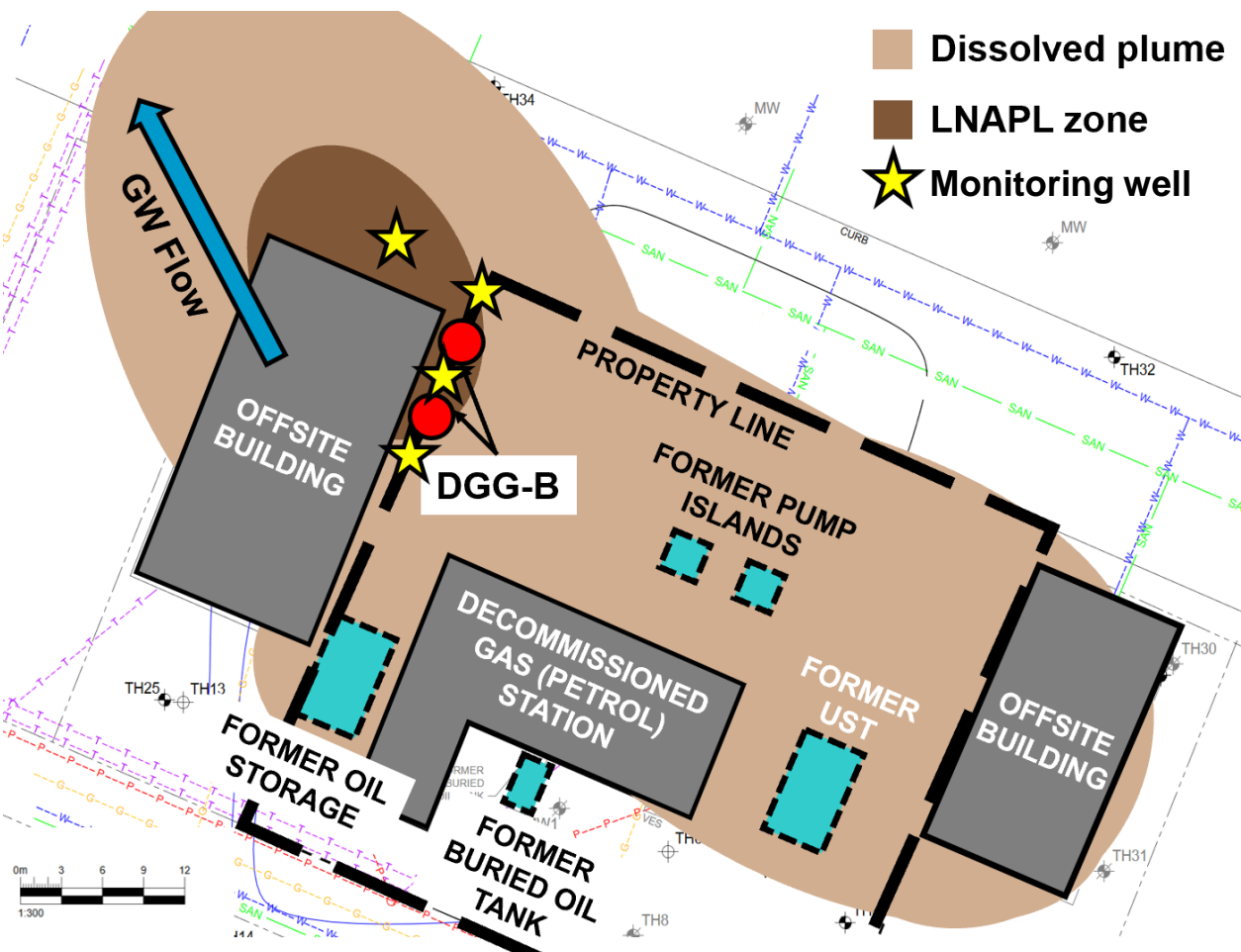
Drilling Rig
and
Injection
Probe

DGG-B

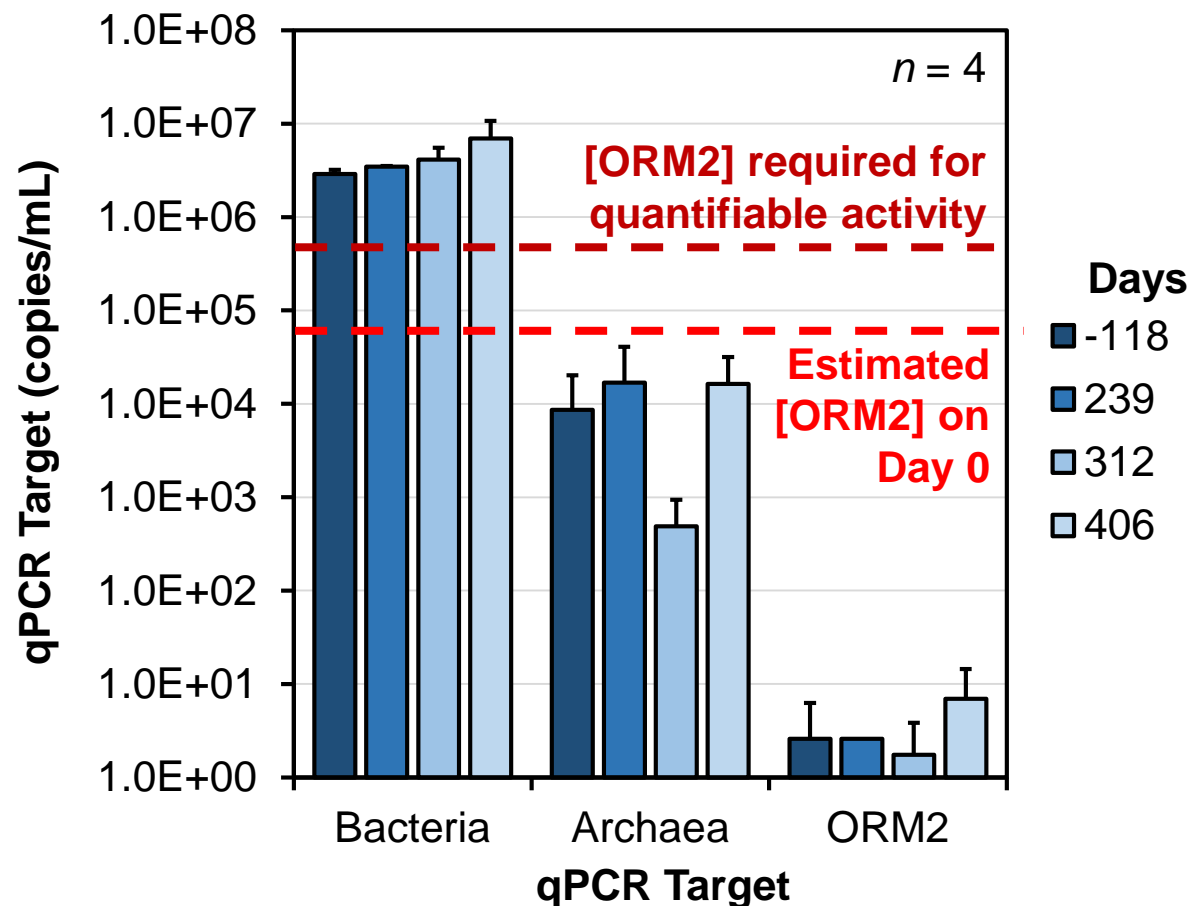
Anaerobic
Water

What about ORM2?

No enrichment of ORM2 has yet to be observed. Perhaps DGG-B did not survive post-injection and/or was poorly dispersed? If cells survived, are they attached to sediments?

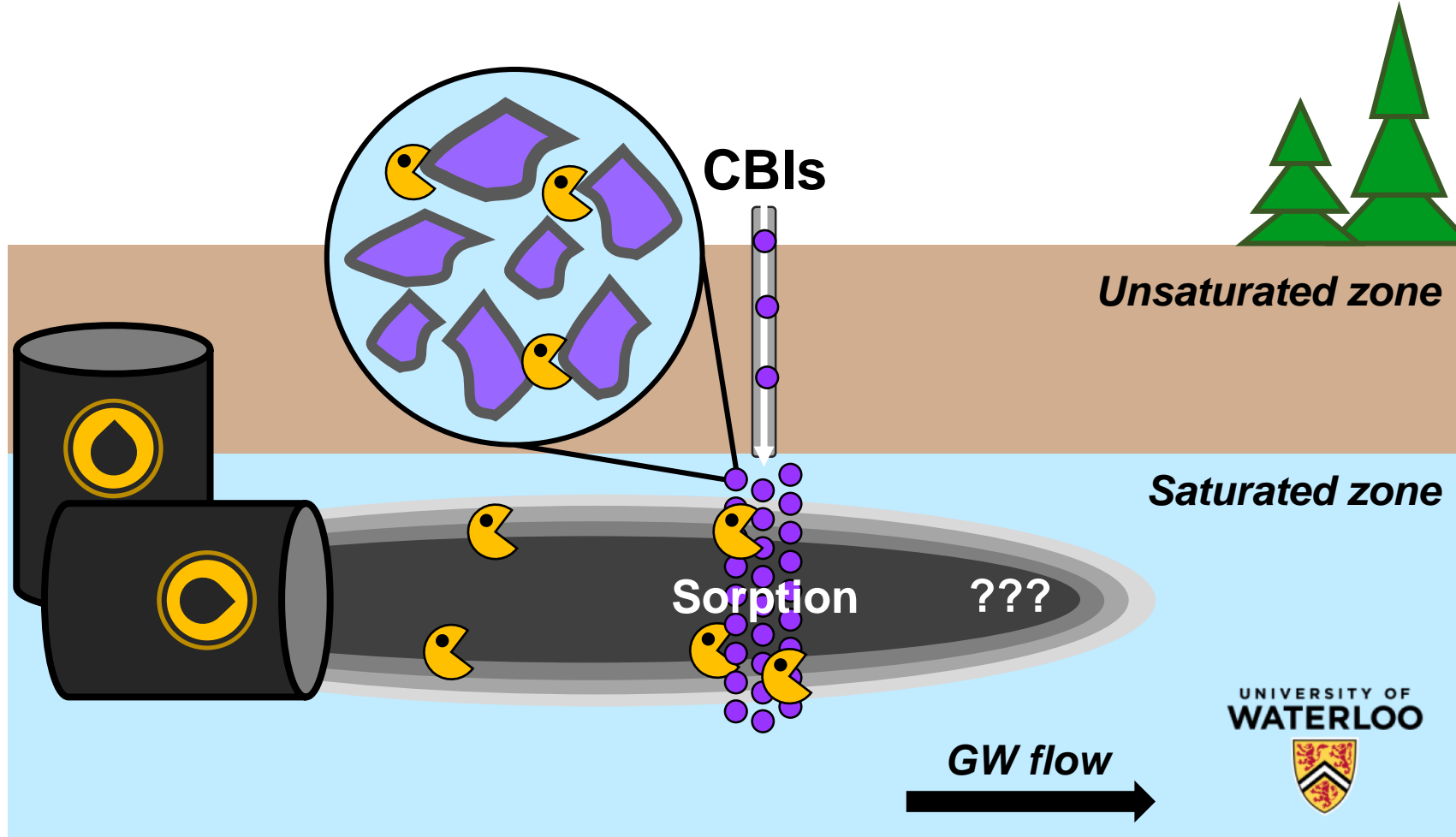


Monitoring Wells Near DGG-B Injection



Can We Improve Bioaugmentation Success?

Strategy #4) Immobilize BTEX on Carbon-Based Injectates (CBIs), encourage localized growth of anaerobic degraders. This technology could be combined with bioaugmentation



Andrea Marrocco



Bill McLaren



Adam Schneider

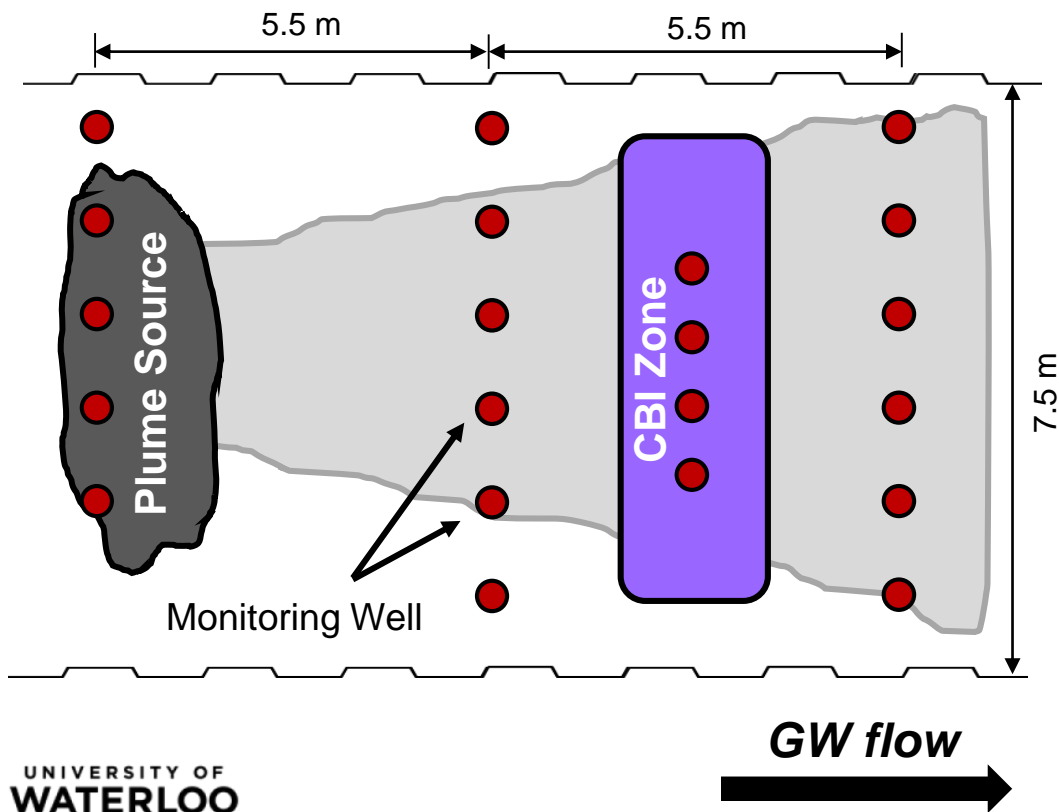


Griselda Diaz de Leon

Can We Improve Bioaugmentation Success?

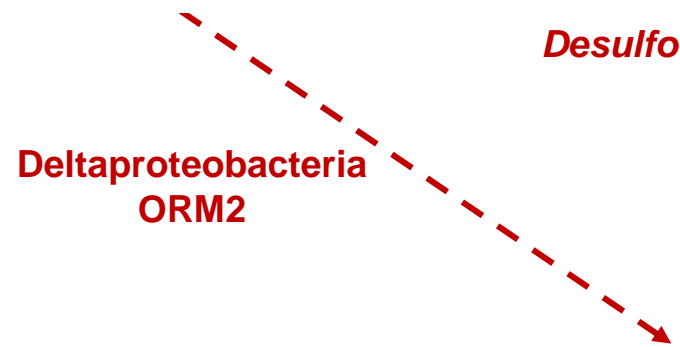
Strategy #4) Immobilize BTEX on Carbon-Based Injectates (CBIs), encourage localized growth of anaerobic degraders. This technology could be combined with bioaugmentation

CBI Field Pilot (Underway)



CBI + DGG-B Microcosm & Column Studies (Underway)





Recyclers

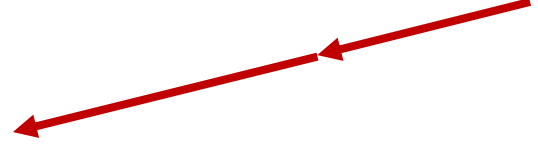
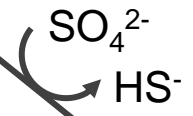


Syntrophs



Sulfate-Reducing Bacteria

Methanogens





Take Home Messages

1. Anaerobic BTEX biodegradation requires high concentrations of active, specialized microbes;
 - 4.3×10^5 ORM2 copies/mL required for benzene
2. These microbes have been scaled up to commercial volumes for field use
3. Our data supports that bioaugmentation can be used to treat BTEX in anaerobic systems;
4. Patience is a virtue





SiREM
Leading Science · Lasting Solutions

DGG-B **DGG-B Anaerobic Benzene Bioaugmentation Culture**

Benzene
Benzene is found naturally in petroleum, and is used extensively in the synthesis of a wide range of materials and chemicals. It is also a frequent groundwater contaminant that is highly toxic and carcinogenic and thus a top priority for remediation efforts.

- Benzene is a specific problem - as it is more toxic and less biodegradable than other TCE (trichloroethylene, ethylbenzene and xylene) compounds (particularly under anaerobic conditions)
- Under anaerobic conditions, benzene is a substantial bottleneck to achieving site cleanup goals.
- Aerobic benzene biodegradation occurs readily; however, injection of sufficient oxygen into the subsurface is often prohibitively expensive. Anaerobic microbial transformation processes offer an attractive remediation alternative and have been successfully applied to chlorinated solvents.
- Why not benzene and other nonchlorinated hydrocarbon compounds?
- Closely related compounds, like toluene, ethylbenzene and xylene, are also readily degraded anaerobically and the microbes that biodegrade these compounds are widely distributed.

The DGG-B benzene-degrading culture is currently available for testing in laboratory and pilot testing.

Contact SiREM for more information on our anaerobic benzene cultures and treatability testing options.

Sandra Dworatzak
(519) 515-0839
sdworatz@siremlab.com

Introducing DGG-B
At the University of Toronto, Elizabeth Edwards' lab has been studying anaerobic degradation of aromatic compounds for decades. A benzene-utilizing, methanogenic culture was derived from an oil refinery site and coupled benzene oxidation with methanogenesis. The culture consistently converts benzene to methane and carbon dioxide (CO₂) at a rate between 0.4 to 1.0 milligrams per liter per day (mg/L/day) and has a double reading conditions. Methanogenic benzene degradation occurs syntrophically by benzene-degrading, fermentative bacteria and methanogens. A *Desulfotomaculum*, designated CR162, has been identified as the key benzene-degrading organism.

Quantitative PCR (qPCR) tools are available to track the abundance of the primary benzene-degrading strain CR162.
qPCR tools have been developed for potential biomarker genes for anaerobic benzene biodegradation. A variety of biomarkers including Gene-Trac® CR162 and S162 can be used to assess the potential benzene-degrading populations that may be present in the environment, either naturally occurring or as a result of biostimulation or bioaugmentation. Contact SiREM for the most current list of applicable biomarkers to test.

Additionally, benzene is attenuating under nitrate electron acceptor conditions. SiREM has a Gene-Trac® test to quantify the presence of *Peptococcus*, the microbes that have been associated with this degradation pathway.

Questions

519-515-0840

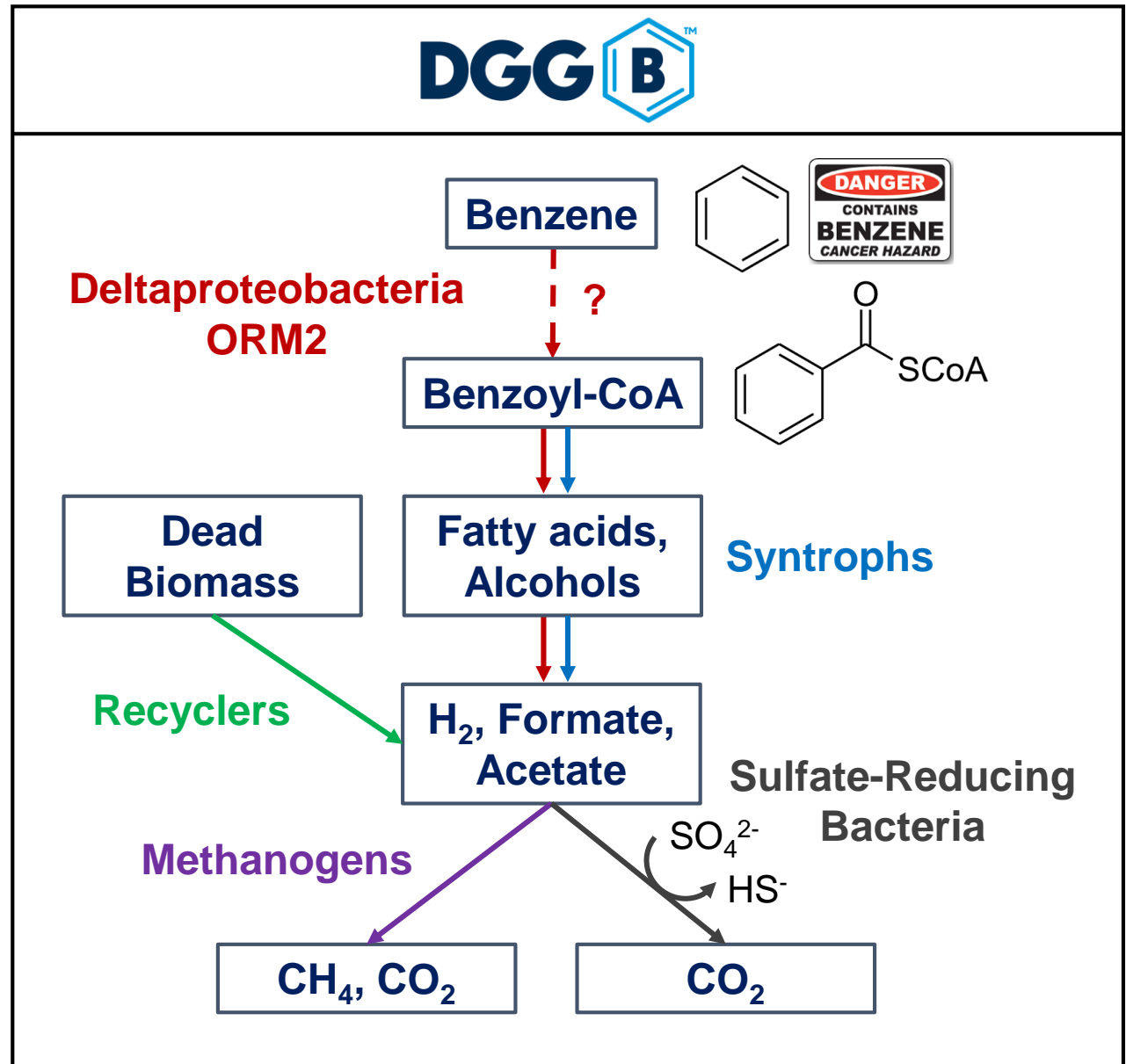
JRRoberts@siremlab.com



<https://www.siremlab.com/advanced-bioaugmentation-cultures/>



Objective: evaluate performance of bioaugmentation for anaerobic benzene treatment



Culture Scale-up & Growth



Lab Treatability Testing



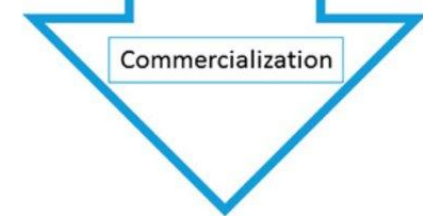
Develop Genomic Monitoring Tools



Regulatory Approval



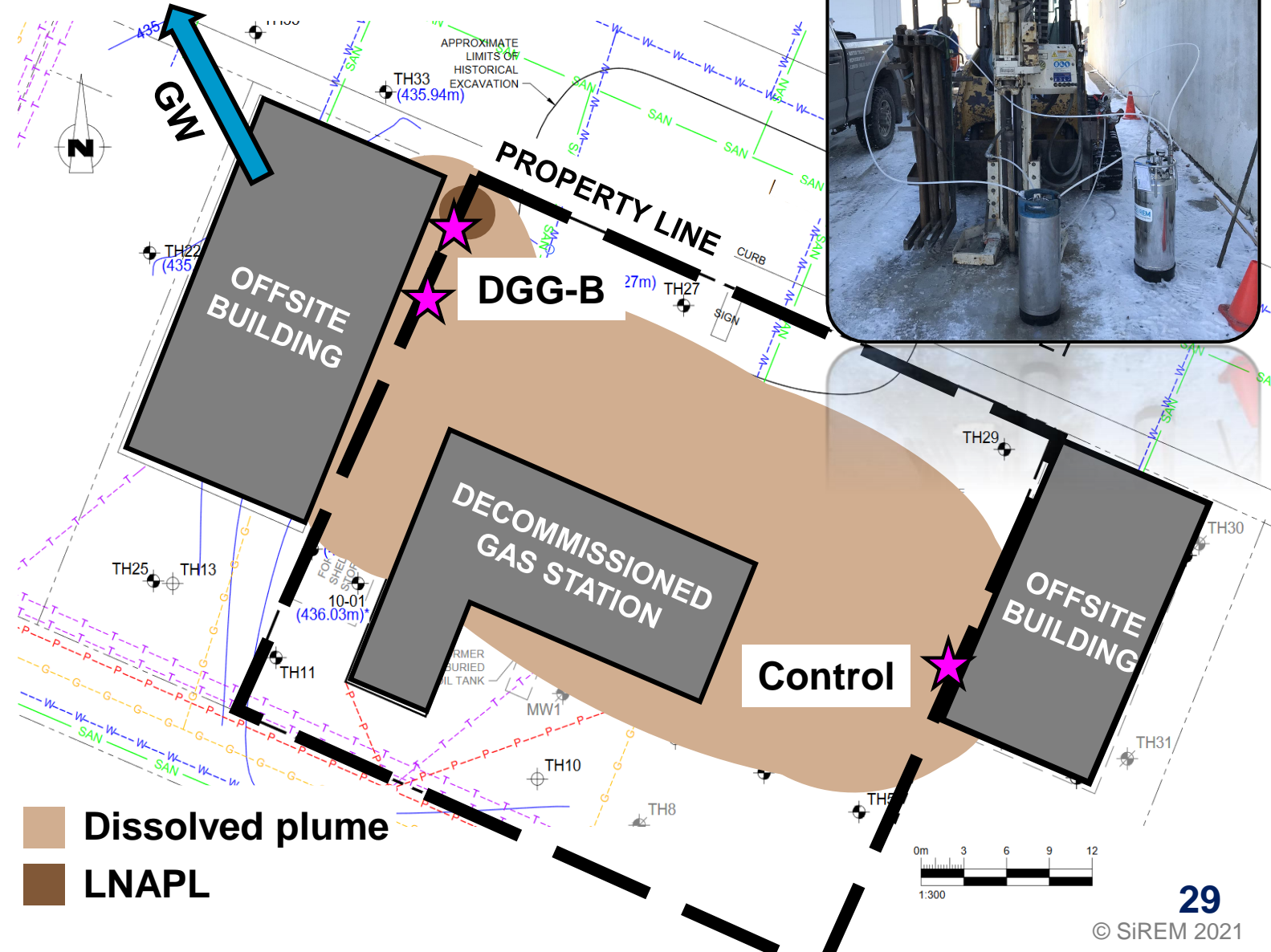
Field Application of Culture



Commercialization

Field Application of DGG-B

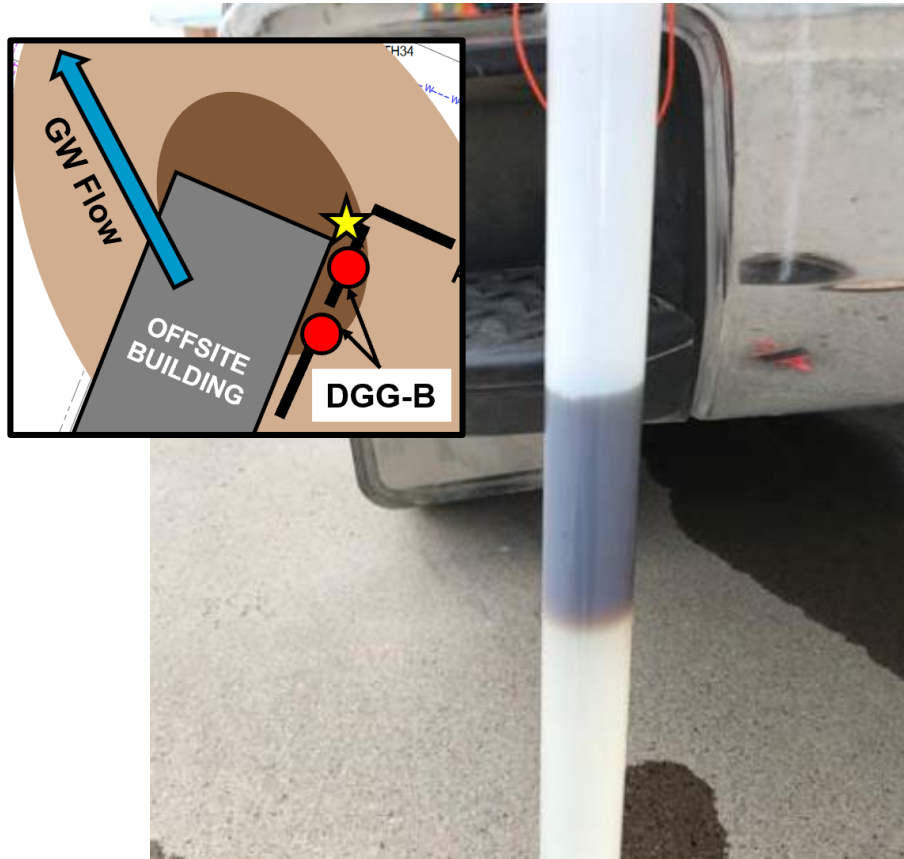
- Decommissioned gasoline (petrol) station with historical BTEX, F1 and F2 contamination
 - Benzene conc. vary ($< 0.01 - 20$ mg/L)
- DGG-B was injected at 2 points (10 L each) near NW corner of property
- A **control well** injected with heat-killed DGG-B (10 L) was established on E edge of property



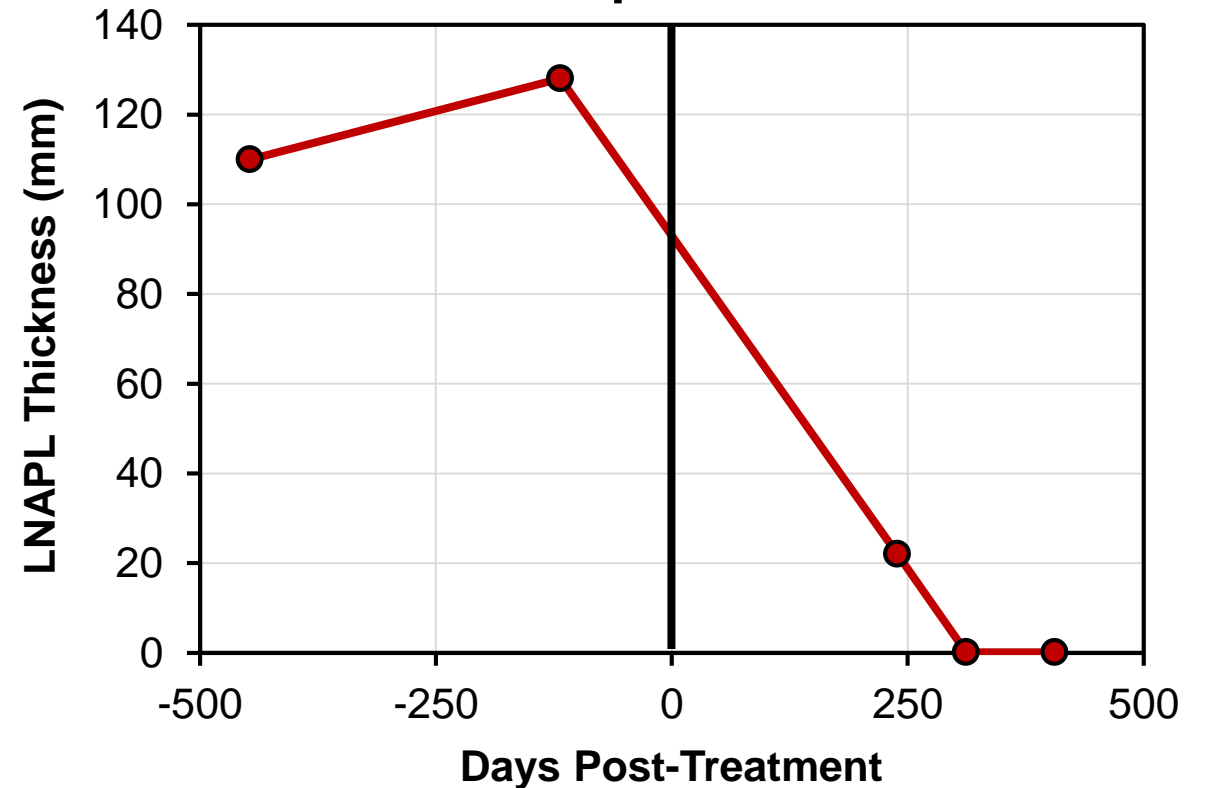
Decrease in LNAPL Thickness?

BTEX biodegradation activity might be masked by LNAPL re-dissolving into the surrounding groundwater. To be verified this year.

Photo of LNAPL layer in Monitoring Well TH28



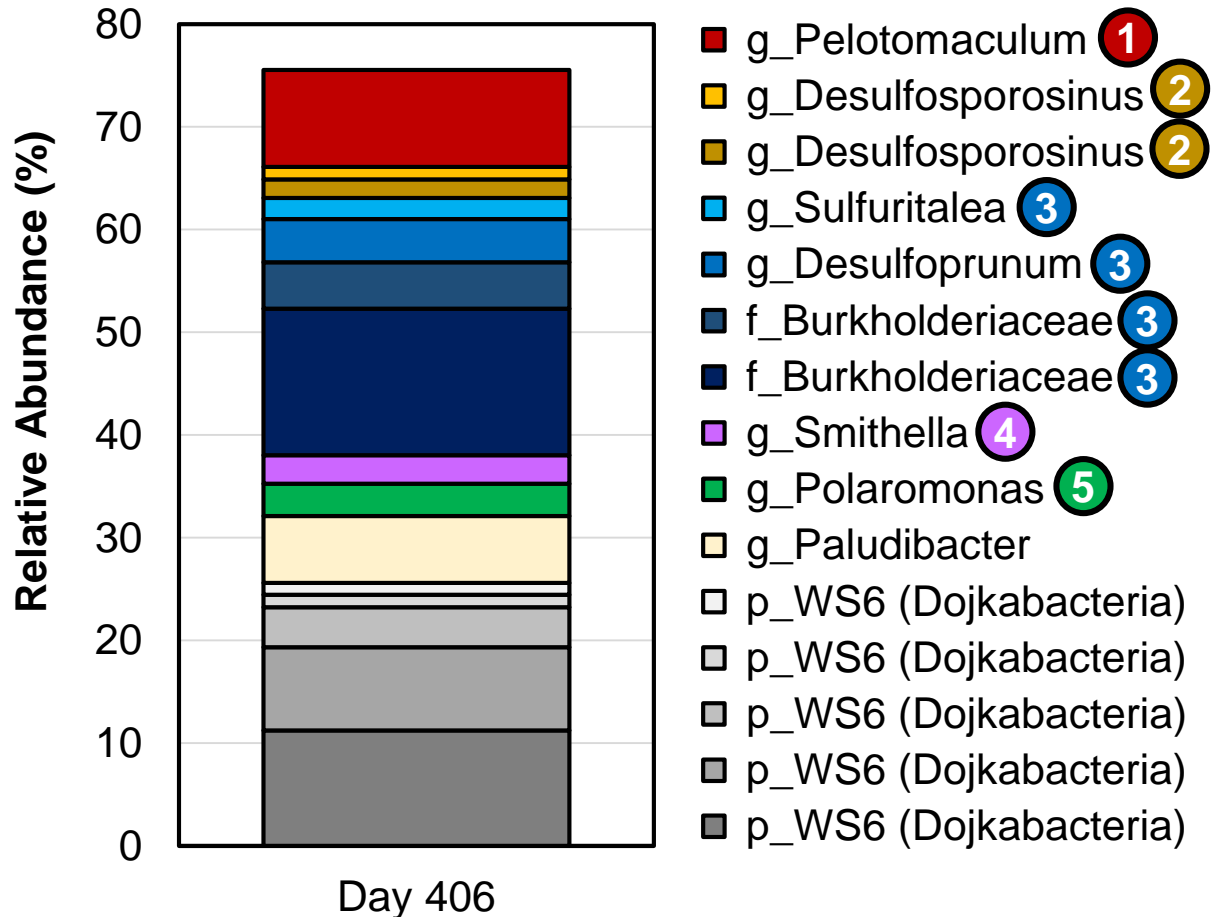
LNAPL Depletion in TH28



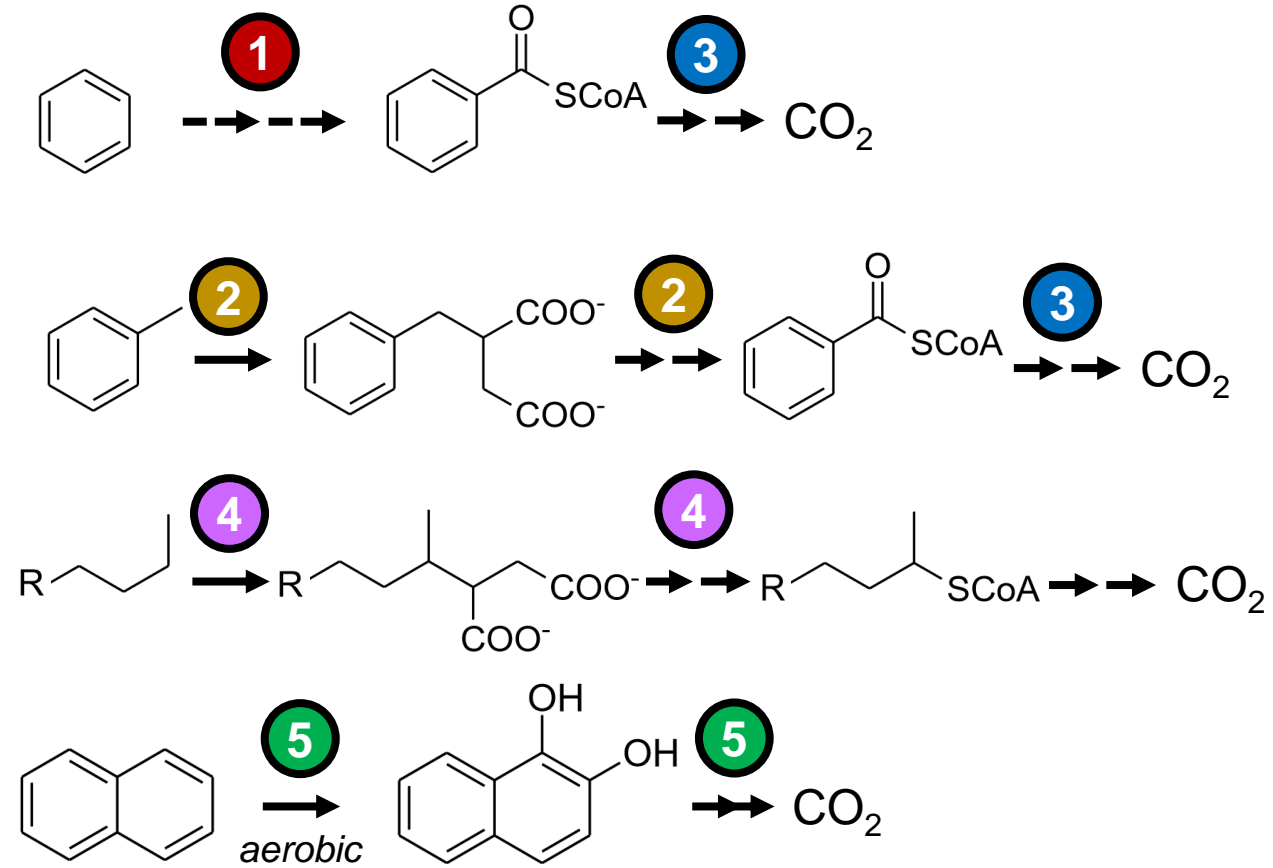
16S rRNA Gene Amplicon Sequencing of TH28

Several putative hydrocarbon degraders were identified, but most appear to be indigenous in origin (i.e., NOT from DGG-B)

Top 15 ASVs



Possible Metabolisms (Based on Taxonomy)



Field Injection Parameters

	Field Pilot (per injection point)	Microcosm Study
Input Parameters		
Concentration of ORM2 in DGG-B Inoculum (copies/mL)	5.2E+07	2.7E+07
Volume of DGG-B added per well (L)	10	2.5E-03
Minimum Dispersed Concentration of ORM2 (DGG-B) at Time of Injection		
Groundwater pore volume (PV, L)	10195	0.2
Number of ORM2 cells inoculated per well ($N_{KB-1} = C_{KB-1} * V_{KB-1}$, copies)	5.2E+11	5.2E+07
Minimum dispersed [ORM2] in groundwater ($C_{PV} = N_{ORM2}/PV$, copies/mL)	5.1E+04	3.4E+05
Expected Lag Time Prior to Onset of Active Biodegradation		
Minimum [ORM2] to achieve quantifiable activity (C_{viable} , copies/mL)	4.3E+05 ^a	
# ORM2 doublings to reach viable concentration [$N_D = \ln(C_{viable}/C_{PV})/\ln(2)$]	6.4	0.7
Estimated time required to achieve one ORM2 population doubling (T_D , days)	50 ^a	
Estimated lag time prior to onset of active degradation ($T_{max\ lag} = T_D * N_D$, days)	154	18
	Lag time (weeks)	Lag time (weeks)
	22.1	2.6

^aFrom Toth et al. 2021 (Environ Sci Technol, *in press*)

Lessons Learned & Take Home Messages

1. Anaerobic BTEX bioremediation requires high concentrations of active, specialized microbes;
 - 10^7 to 10^8+ copies/L required for benzene
2. Even with bioaugmentation, it is challenging to get/keep their numbers up;
 - Re-bioaugmentation occurred in Summer 2021
 - Additional measures will be included to understand what is happening to DGG-B post-bioaugmentation
 - What else can we do to improve bioaugmentation success?
3. Patience is a virtue – *this study will take years*

Thank You for Your Attention



Contact Us!



Courtney Toth, PhD
(PDF/Manager, UofT)
courtney.toth@utoronto.ca



Sandra Dworatzek, MSc
(Receptor Lead, SiREM)
SDworatzek@siremlab.com



Elizabeth Edwards, PhD
(Academic Lead, UofT)
elizabeth.edwards@utoronto.ca