Development of an Amendment Recipe and Identification of Benzene Degraders for Anaerobic Benzene Bioremediation

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## Outline

Introduction

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**3** Materials and Methods

4 Results and Discussion





# 1 Introduction

- Petroleum hydrocarbons (PHC) release into soil and groundwater occur at every stage of oil extraction, refinement, storage, transportation, and disposal.
- Benzene contamination induced by PHC release is a major concern.
  - Highly water soluble (i.e., solubility of 1,791 mg/L at 25°C)
  - Volatile
  - Group A human carcinogen
  - A maximum acceptable concentration of 5 µg/L for benzene in drinking water



# 1 Introduction

- Benzene is readily biodegradable under aerobic conditions
- Anaerobic subsurface environmental development
  - Fast oxygen consumption rate
  - Slow oxygen supply rate
- Anaerobic reducing conditions typically predominate
- Anaerobic intrinsic benzene biodegradation
  - Occurs very slowly
  - Incomplete
  - Long lag time



# 1 Introduction

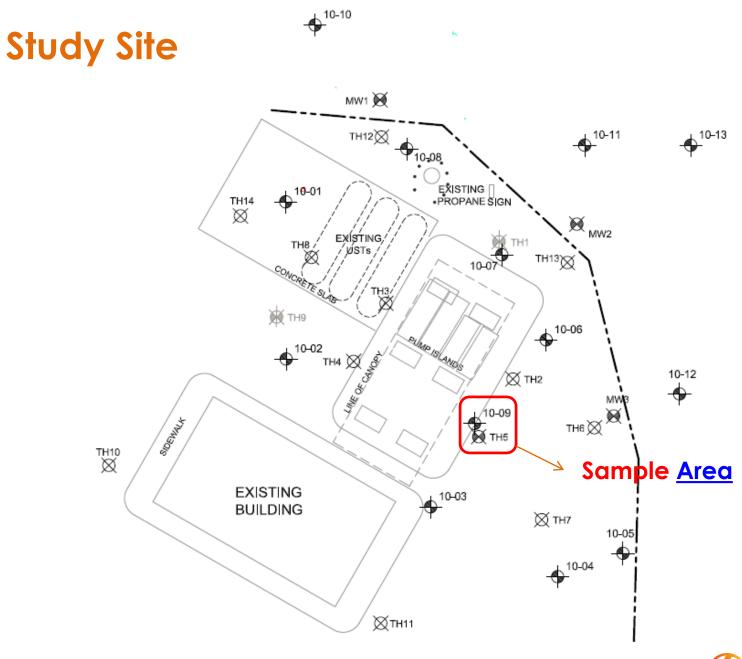
- Enhanced anaerobic bioremediation (EAB) is a practical and cost effective PHC remediation method
  - Biostimulation: addition of nutrients and electron acceptors
  - Bioaugmentation: addition of microorganisms
- Success of biostimulation
  - An effective configuration of nutrients and electron
    acceptors
  - A community structure of dominant microbes
- Understanding the indigenous microbial community is also critical for bioaugmentation



# 2 Objectives

- To establish optimal chemical composition of nutrients and electron acceptors
- To identify dominant benzene-degrading consortium using molecular approaches







#### **Soil Characteristics**

Parameters	Value
Benzene (mg/kg)	<u>50</u>
Toluene (mg/kg)	10.3
Ethylbenzene (mg/kg)	251
Xylenes (mg/kg)	1080
Petroleum Hydrocarbon F1 (mg/kg)	9680
Petroleum Hydrocarbon F2 (mg/kg)	2990
Petroleum Hydrocarbon F3 (mg/kg)	120



#### **Groundwater Characteristics**

Parameters	Value
Temperature (°C)	11.13
рН	6.59
Electrical Conductivity (µs/cm)	8.13
Oxidation Reduction Potential (mV)	- 40
Dissolved Oxygen (mg/L)	0.56
Nitrate (mg/L)	1.24
Phosphate (mg/L)	0.02
Sulphate (mg/L)	14.8
Benzene (mg/L)	<u>10.7</u>
Toluene (mg/L)	1.12
Ethylbenzene (mg/L)	3.18
Xylenes (mg/L)	3.1
Petroleum Hydrocarbon F1 (mg/L)	15
Petroleum Hydrocarbon F2 (mg/L)	4.72
Petroleum Hydrocarbon F3 (mg/L)	0.051



#### **Amendment Recipes**

Recipe	Media Composition (each litre)	Benzene Concentration (mg/L)
NP	3.5 g NaNO <sub>3</sub> , 0.3 g K <sub>2</sub> HPO <sub>4</sub> , 1.49 mg FeCl <sub>2</sub> ·4H <sub>2</sub> O	<u>25</u>
Coates	4.0 g Na <sub>2</sub> SO <sub>4</sub> , 1.36 g <u>sodium acetate</u> , <u>20 g NaC</u> I, 0.5 g KCI, 0.2 g KH <sub>2</sub> PO <sub>4</sub> , 0.25 g NH <sub>4</sub> CI, 0.15 g CaCl <sub>2</sub> ·2H <sub>2</sub> O, 3 g MgCl <sub>2</sub> ·H <sub>2</sub> O, 1.49 mg FeCl <sub>2</sub> ·4H <sub>2</sub> O	25
Kazumi	2.84 g Na <sub>2</sub> SO <sub>4</sub> , 1.3 g KCl, 0.2 g KH <sub>2</sub> PO <sub>4</sub> , 23 g NaCl, 0.5 g NH <sub>4</sub> Cl, 0.1 g CaCl <sub>2</sub> ·2H <sub>2</sub> O, 1 g MgCl <sub>2</sub> ·H <sub>2</sub> O, 2.5 g NaHCO <sub>3</sub> , 1.49 mg FeCl <sub>2</sub> ·4H <sub>2</sub> O	17
SA	2.7 g K <sub>2</sub> SO <sub>4</sub> , 0.5 g KNO <sub>3</sub> , 1 g KH <sub>2</sub> PO <sub>4</sub> , 0.5 g NH <sub>4</sub> Cl, 0.14 g CaCl <sub>2</sub> .2H <sub>2</sub> O, 1 g MgCl <sub>2</sub> .6H <sub>2</sub> O, 2.5 g NaHCO <sub>3</sub> , 2 mg FeCl <sub>2</sub> .4H <sub>2</sub> O	17
SA+T	10 mg toluene 2.7 g K <sub>2</sub> SO <sub>4</sub> , 0.5 g KNO <sub>3</sub> , 1 g KH <sub>2</sub> PO <sub>4</sub> , 0.5 g NH <sub>4</sub> Cl, 0.14 g CaCl <sub>2</sub> .2H <sub>2</sub> O, 1 g MgCl <sub>2</sub> .6H <sub>2</sub> O, 2.5 g NaHCO <sub>3</sub> , 2 mg FeCl <sub>2</sub> .4H <sub>2</sub> O	17
SA+BP	25 mg benzoate, 35 mg phenol 2.7 g K <sub>2</sub> SO <sub>4</sub> , 0.5 g KNO <sub>3</sub> , 1 g KH <sub>2</sub> PO <sub>4</sub> , 0.5 g NH <sub>4</sub> Cl, 0.14 g CaCl <sub>2</sub> .2H <sub>2</sub> O, 1 g MgCl <sub>2</sub> .6H <sub>2</sub> O, 2.5 g NaHCO <sub>3</sub> , 2 mg FeCl <sub>2</sub> .4H <sub>2</sub> O	17



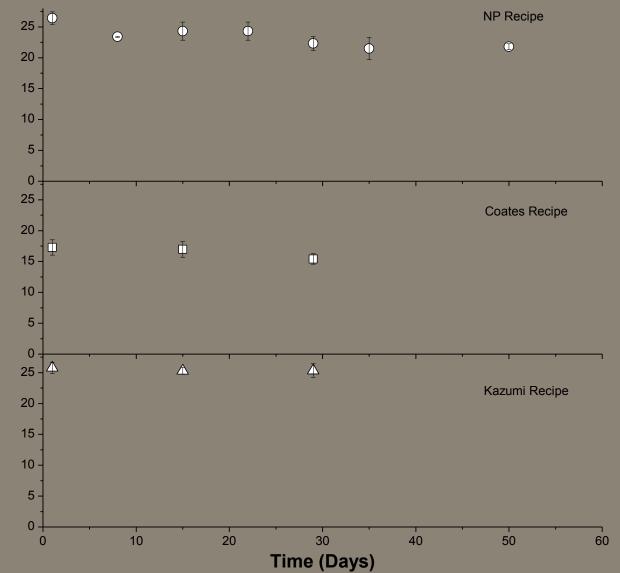
#### **Microbial Analysis**

- BioSep BioTrap pre-amended with [<sup>13</sup>C] benzene
- Soil, groundwater, BioSep BioTrap was incubated with recipe SA+T
- Analysis of [<sup>13</sup>C] of dissolved inorganic carbon
- Analysis of fatty acid methyl esters (FAMEs)
- Analysis of real-time quantitative polymer chain reaction (qPCR)
- Analyses of denaturing gradient gel electrophoresis (DGGE) and 16S rRNA



## 4 **Results and Discussion**

#### **Recipes NP, Coates, and Kazumi**



**3enzene Concentrations (mg/L**)

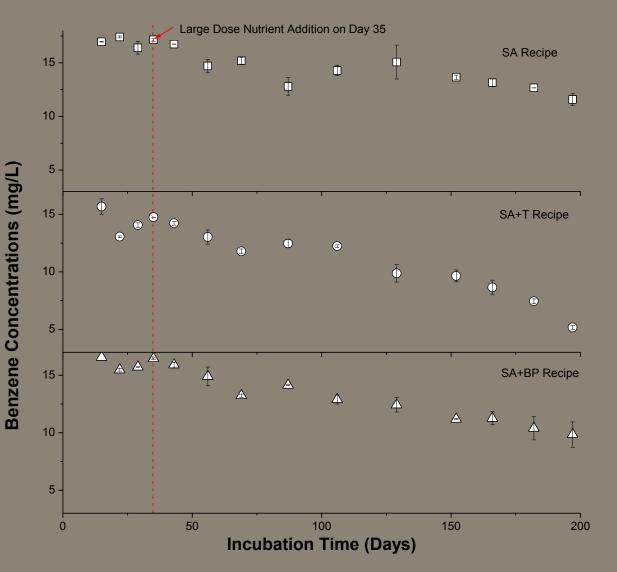
Benzene was degraded only slightly in microcosms incubated with Recipe NP, Coates, and Kazumi.

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Salinity, high initial benzene concentration, <u>alternative carbon</u> source, and an extended lag time may result in the low benzene degradation rate.



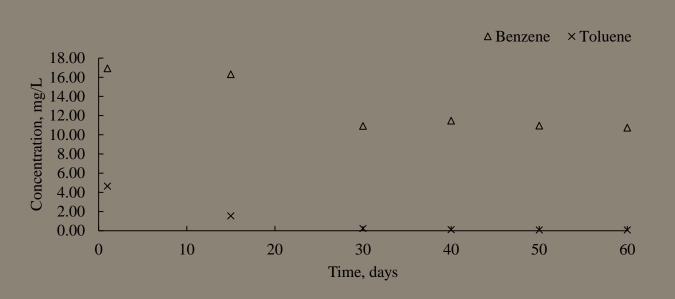
## 4 Results and Discussion Recipes SA, SA+T, SA+BP



- The lag time was overcome by the large-dose addition of the amendments.
  - Recipe SA+T with toluene amendment achieved the highest benzene degradation rate.



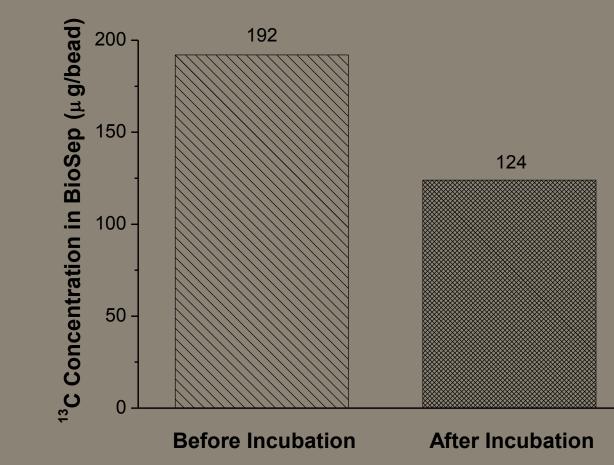
## 4 Results and Discussion Toluene's Role on Benzene Biodegradation



- Both toluene and benzene were degraded effectively in a rapid manner during the first 30 days.
- After 30 days, benzene degradation stopped with the toluene depletion.



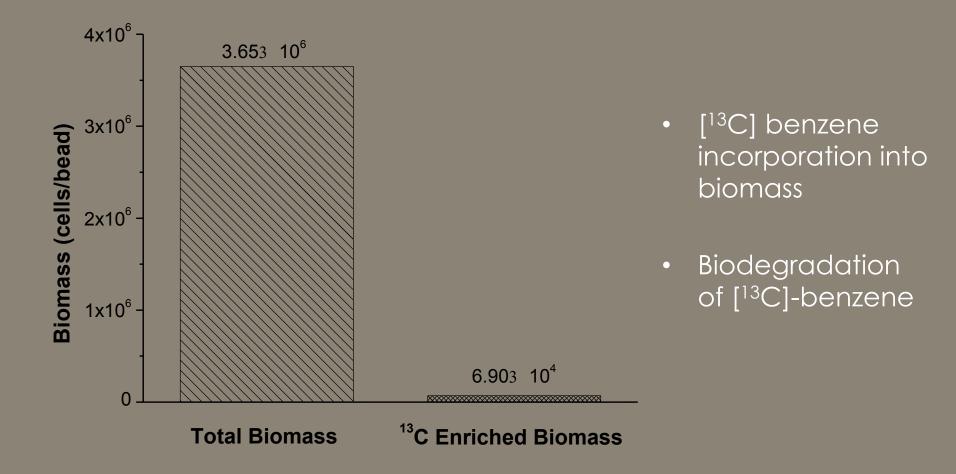
## 4 Results and Discussion [<sup>13</sup>C] Concentrations on BioSep Beads



- Degradation rate of [<sup>13</sup>C] benzene was calculated to be 35.42%.
- BioSep beads could
  support microorganism
  growth.



### 4 Results and Discussion Total and [<sup>13</sup>C]-enriched biomass





# 4 Results and Discussion

Community Structure	% of Total PLFA
Proteobacteria (Monos)	63.1
General (Nsats)	32.9
Firmucutes (TerBrSats)	2.8
Anaerobic metal reducers (BrMonos)	1.1
Actinomycetes (MidBrSats)	0.2
Eukaryotes (Polyenoics)	0.0



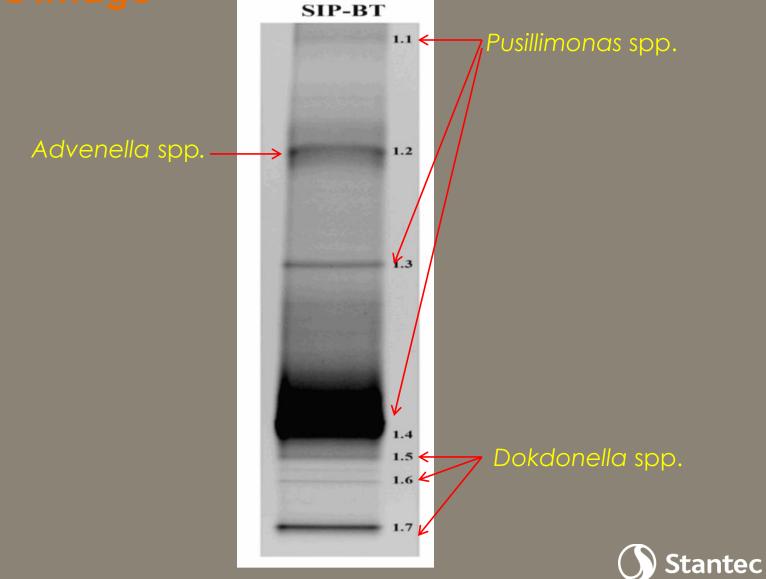
## 4 Results and Discussion qPCR Analysis

Functional Genes	Gene Copy Numbers	
Benzene carboxylase (abcA)	<50	
Benzylsuccinate synthase (bssA)	<50	
nirS	2.01×10 <sup>6</sup>	
nirK	3.05×10 <sup>7</sup>	
Phylogenetic Group		
APS	5.13×10 <sup>2</sup>	



# 4 **Results and Discussion**

#### **DGGE Image**



# 4 **Results and Discussion**

#### **16S rRNA Analysis**

Band	Similar Genus	Similarity Index	Affiliation	GenBank Accession Number
1.1	Pusillimonas spp.	0.926	Betaproteobacteria; Alcaligenaceae	FN667020.1
1.2	Advenella spp.	0.880	Betaproteobacteria; Alcaligenaceae	KC464861.1; KC207092.1; JQ799008.1
1.3	Pusillimonas spp.	0.990	Betaproteobacteria; Alcaligenaceae	KC464818.1; HQ326782.1; FN667020.1
1.4	Pusillimonas spp.	0.923	Betaproteobacteria; Alcaligenaceae	FM956659; FJ791048; GQ246953
1.5	Dokdonella spp.	0.980	Gammaproteobacteria; Xanthomonadaceae	JQ726695.1; JQ726692.1; JQ726691.1
1.6	Dokdonella spp.	0.970	Gammaproteobacteria; Xanthomonadaceae	JQ726695.1; JQ726692.1; JQ726691.1
1.7	Dokdonella spp.	0.876	Gammaproteobacteria; Xanthomonadaceae	AY921834.1



# 5 Lessons Leaned / Conclusions

- Salinity and a high initial concentration of benzene were detrimental for benzene biodegradation.
- A large dose of amendment can shorten the lag time for benzene biodegradation.
- Toluene was an essential co-substance for promoting benzene biodegradation.
- Incorporation of <sup>13</sup>C-benzene into microbes provided direct evidence for benzene biodegradation.
- Dominant mechanism for benzene biodegradation was nitrate reduction in this study.
- Dokdonella spp., Pusillimonas spp., and Advenella spp. were predominant in the microbial community and involved in anaerobic benzene bioremediation.



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Thank you

